

VOLUME 6 ISSUE 1

JUNE 2012



**University of Duhok
College of Medicine**

Duhok Medical Journal

The Official Journal of Duhok College of Medicine

Duhok University Press

ISSN: 2071-7326

This page is left intentionally

Duhok Med J

EDITORIAL BOARD

PATRON

Dr. ARIF Y. BALATAY, MBChB, PhD (Ophthalmology)

Dean, Faculty of Medical Sciences, University of Duhok

EDITOR-IN-CHIEF

Prof. SAMIM A. AL-DABBAGH, MBChB, DTM&H, D. Phil

Head, Department of Family and Community Medicine, Duhok College of Medicine

MEMBER

Prof. DHIA J. AL-TIMIMI, BSc (pharm), Mphil, PhD

Head, Department of Clinical Biochemistry, Duhok College of Medicine

MEMBER

Prof. NASIR A. AL-ALLAWI, MBChB, MSc, PhD

Head, Department of Pathology, Duhok College of Medicine

MEMBER

Dr. FARHAD K. SULAYVANI, MBChB, CABS, FRCS

Assistant professor, Department of Surgery, Duhok College of Medicine

MEMBER

Dr. MAIDA Y. SHAMDEEN, MBChB, MRCOG, RECOG

Assistant professor, Department of Obstetrics and Gynecology, Duhok College of
Medicine

MEMBER

Dr. MOHAMMED T. RASOOL, MBChB, FRCPG, FRCP (London)

**Assistant professor, Head, Department of Internal Medicine, Duhok College of
Medicine**

MEMBER

Dr. ABDULGHAFOOR S. ABDULKAREEM, MBChB, FICMS

Assistant professor of Urology, Department of Surgery, Duhok College of Medicine

EDITORIAL ASSISTANT

Dr. ABDULLA J. RAJAB, MBChB, MPH

Director of Department of Continuing Medical Education, Duhok Directorate of Health

DESIGNER

Dr. HUSHYAR M. SULAIMAN, MBChB, MSc, MHS (Health Policy)

Department of Continuing Medical Education, Duhok Directorate of Health

Submission of Manuscript:

Manuscripts should be submitted to:

The Editor,
Duhok Medical Journal,
Duhok College of Medicine,

Post address: Nakhoshkhana Road 9, 1014, AM, Duhok, Iraq.

Telephone No.: 00964-62-7224268 EXT 115

E-mail: dmj.med.uod@gmail.com

Electronic submission of articles is also accepted

Duhok Med J

ADVISORY BOARD

Prof. GAZI ZIBARI, MD, FACS, FICS

Director of W.K./L.S.U. Regional Transplant Program, Louisiana, USA

Prof. AHMAD MB. AL-KAJAJEI, MBChB, DTM&H, PhD, MFCM

Head, Department of Public Health, Jordanian College of Medical Sciences

Prof. FAYSIL A. ALNASIR, FPC, FRCGP, MICGP, PhD

Vice President, Arabian Gulf University, Bahrain

Dr. ASAD A. ZOMA FRCP, FRCPG, FACR

**Consultant Physician in Rheumatology and Senior Clinical Lecturer
Lanarkshire Health Board and Glasgow University, Scotland, United Kingdom**

Dr. NADA J. AL-WARD, MBChB, MFCM

Public Health Specialist, WHO, Geneva

Dr. CHRISTINE M. EVANS, MBChB, MD Ed, FRCS, FRCS Ed

Urologist, North Wales, United Kingdom

Dr. FARHAD U. HUWEZ, MBChB, PhD, MRCPI, FRCP, FRCPG

**Consultant Physician / Lead Physician of Stroke Services, Basildon & Thurrock NHS
Trust, Basildon Hospital, United Kingdom**

Dr. ABDULBAGHI AHMAD, MD, PhD

**Consultant Child Psychiatrist and Director of Studies, Department of Neuroscience,
Child and Adolescence Psychiatry, Uppsala University Hospital, Sweden**

This page is left intentionally

Duhok Med J

INSTRUCTIONS FOR AUTHORS

Aims and Scope Duhok Medical Journal is a peer reviewed journal issued bi – annually by Duhok College of Medicine. Scientific and clinical researches are the main issues. The journal also publishes short articles, letters to editors, review articles and case reports.

General The Duhok Medical Journal is a signatory journal to the uniform requirement for manuscripts submitted to biomedical journals, February 2006 [updated 2009] (<http://www.icmje.org>).

To present your original work for consideration three manuscript copies written in English together with Kurdish and Arabic abstracts should be submitted to the editor. All authors are required to provide the manuscript on a CD labeled with the name and title of the paper.

Preparation of the manuscript The manuscript should be typed double spaced as normal text on one side of the paper in single column format, font size 14 pt, paper type A4, 1" margin at each side and each of the following sections should begin on a new page in the following sequence:

- 1- **Title page**; should include the following: title, font size 16 pt, each author's full name, academic degree(s), scientific title (if available), institutional affiliation, full contact information including emails. If there are more than one author, article should include author to whom correspondence should be addressed including the scientific title (if available), institution affiliation, address, email, telephone.
- 2- **Structured abstract**; of no more than 250 words including background and objectives, methods, results, and conclusions.
3 – 10 keywords or phrases should be put at the end of each abstract (Printed in bold font; size 12 pt).
- 3- **Body of the text**; structured in an IMRAD style;
(Introduction, Methods, Results and Discussion).
- 4- **Acknowledgment** (if any.)
- 5- **References.**
- 6- **Tables with legends.**
- 7- **Illustrations with legends.**
- 8- **Structured Kurdish abstract including title in Kurdish.**
پێشهکی و ئارمانج، رێکێن فهکولینی، نه‌نجام، ده‌ره‌نجام
- 9- **Structured Arabic abstract including title in Arabic.**
خلفية و اهداف البحث، طرق البحث، النتائج، الاستنتاجات

Tables Each table must be typed on separate page and should follow the reference list. All the tables must be numbered consecutively in the order of their first citation in the text. Supply a brief title for each on top and place explanatory matter in foot notes not in the heading (if needed). Tables should be simple and not duplicated in the text. Percentages are included with numbers in the same cells but in brackets.

Illustrations Graphs, line drawing, photographs, printed x rays and other illustrations are accepted only if they add to the evidence of the text. They should be of a high quality and suitable for reproduction. They should be numbered consecutively according to the order in which they have been first cited in the text. Supply a brief title beneath each illustration. Graphs should have white background; should be colored and non 3-dimensional figure; and should have labels for X and Y axis.

Numbers and Units Measurements of length, height, weight and volume should be reported in metric units. Temperature in degrees Celsius, blood pressure should be expressed in mmHg and all hematologic and clinical chemistry measurements in SI units.

Abbreviations should be defined on first use and then applied consistently throughout the article. Avoid abbreviations in the title and abstract.

References should be numbered both in text and in the list of references in the order in which they appear in the text. The punctuation of the Vancouver style should be followed; if the original reference is not verified by the author, it should be given in the list of references followed by (cited by) and the paper it was referring to. The titles of journals should be abbreviated according to the style used in Index Medicus. This can be obtained from website (<http://www.nlm.nih.gov/>). The author is responsible for the accuracy of references. The following are examples of the three most common types of citations:

The article citation: if six authors or fewer list all; if seven or more authors list the first six and then add "et al":

1- Nuwayhid IA, Yamout B, Azar G, Kambris MA. Narghile (hubble bubble) smoking, low birth weight, and other pregnancy outcomes. *Am J Epidemiol.* 1998;148(4):375-83.

Book citation, noting chapter and authors:

2- Arevalo JA, Nesbitt TS. Medical problems during pregnancy. In: Taylor RB, editor. *Family medicine: principles and practice.* 6th ed. New York: Springer – Verlag; 2003. p. 109-16.

Electronic source:

3- Garfinkel PE, Lin E, Goering P. Should amenorrhoea be necessary for the diagnosis of anorexia nervosa? *Br J Psych [Internet].* 1996 [cited 1999 Aug 17];168(4):500-6. Available from: URL:<http://biomed.niss.ac.uk>

Authorship and consent form All authors must give signed consent (Form No.1- Submission Form), which should accompany the manuscript. The letter should say "this manuscript is an unpublished work, which is not under consideration elsewhere in the record. Authors are requested to state an approximate estimate of their contribution in the study, sign the form and send it with the manuscript.

Authors must declare if they have any competing interests in the study and to specify any funds given to conduct the study.

Ethical considerations When experiments on humans are being reported the whole work in the manuscript should conform to the ethical standards of the responsible committee on human experimentation.

Submission of manuscript

Manuscripts should be submitted to:

The Editor,

Duhok Medical Journal,

Duhok College of Medicine,

Post address: Nakhoshkhana Road 9, 1014, AM, Duhok, Iraq.

Telephone no.: 00964-62-7224268 EXT 115

E-mail: dmj.med.uod@gmail.com

Electronic submission of articles is also accepted

N.B.

* Accepted manuscripts may be altered by the editorial board of Duhok Medical Journal to conform to details of the journal publication style.

** The Editorial Board of Duhok Medical Journal accepts no responsibility for statement made by authors in articles published by the journal.

Duhok Med J

CONTENTS

XMN I POLYMORPHISM IN B-THALASSEMIC PATIENTS IN THE DUHOK REGION –IRAQ

NASIR AL-ALLAWI, FARIDA F. NEIRWAY, DILAN JASSIM, SHARAZA Q. OMER, SANA D. JALAL, RAJI D. MARKOUS 1-7

INCIDENCE AND ANTIBIOTIC SUSCEPTIBILITY PATTERNS OF PSEUDOMONAS AERUGINOSA IN BURNS INFECTIONS IN DUHOK CITY

NAJIM A. YASSIN, JANAN M. SALIH, BLIND H. ABDULLA 8-16

BIOFILM FORMATION BY METHICILLIN RESISTANT STAPHYLOCOCCUS AUREUS (MRSA) WITHIN HOSPITAL AND COMMUNITY ACQUIRED URINARY TRACT INFECTIONS

AZA B. TAHA, SABRIA M. SAID AL-SALIHI 17-27

THE EFFECT OF HELICOBACTER PYLORI ERADICATION THERAPY ON PLATELET COUNT IN IDIOPATHIC THROMBOCYTOPENIA: A PILOT STUDY

ASWAD AL-OBEIDY, AHMED Y. ELMESHEDANY 28-33

METHYLENETETRAHYDROFOLATE REDUCTASE (C677T) MUTATION IN HEALTHY INDIVIDUALS IN ERBIL - IRAQ

FARIDA FA. NERWEYI 34-42

CHILDHOOD HENOCCH-SCHÖNLEIN PURPURA IN KURDISH POPULATION OF DUHOK CITY

KHALID N. ABDURRAHMAN, DELAWER B. MIKAEL 43-53

CROHN'S DISEASE IN AN INFANT PRESENTED WITH PERFORATION OF THE COLON: A CASE REPORT

NADIR A. GARJEES, RAFIL T. YAQO, MUHAMMED H. ALDABAKH 54-58

AMIODARONE- INDUCED THYROIDITIS IN A POST-CARDIAC TRANSPLANT PATIENT – CASE REPORT

HATEM AL-FARHAN, AFTAB A. SIDDIQUI, YASSER W. SHAREF, ADIL F. AL LAWATI 59-63

This page is left intentionally

XMN I POLYMORPHISM IN β -THALASSEMIC PATIENTS IN THE DUHOK REGION –IRAQ

NASIR AL-ALLAWI, MBChB, PhD*

FARIDA F. NEIRWAY, BSc, PhD**

DILAN JASSIM, BSc***

SHARAZA Q. OMER, MBChB****

SANA D. JALAL, MBChB, FIBMS*****

RAJI D. MARKOUS, MBChB, HDCH*****

Submitted 14 Nov 2011; accepted 5 Jun 2012

ABSTRACT

Background and objectives Several genetic modifiers have been implicated in phenotypic variations in β -thalassemia(thal) syndromes. Among the frequently implicated ones in the Eastern Mediterranean is Xmn I polymorphism. No study has addressed the relative contribution of this polymorphism to the phenotypic variation of Iraqi β -thalassemia patients.

Methods A total of 107 symptomatic β -thal patients (52 thal Intermedia and 55 thal major) were enrolled. Their clinical records were reviewed. All had full blood counts as well as HbA2 and F quantitation by high performance liquid chromatography performed. The presence of Xmn I polymorphism was documented by PCR-RFLP based method.

Results The homozygous XmnI (+/+) status was found in 19.2% and 1.8% of the thal intermedia and major patients respectively ($p=0.0028$). Heterozygous state Xmn I (+/-) was found in 32.7 and 21.8% respectively. Overall and among the 107 patients enrolled the (+/+), (+/-) and the (-/-) were associated with median annual transfusion rates of 0, 3.5 and 11 respectively, with significantly lower requirements of those with the (+/+) and (+/-) when compared to non-carriers ($p=.004$ and 0.02 respectively). The highest Hb F levels were found among those with the (+/+) and the least in the (-/-), a finding which was significant ($p=0.029$).

Conclusions Xmn I polymorphism is an important genetic modulator of severity of β -thal among Iraqi Kurds, and is associated with higher Hb F levels than non-carriers. Further genetic modulators need to be investigated to determine their relative contributions to the phenotypic variation of this important genetic disease.

Duhok Med J 2012;6(1): 1-7.

Key words: B-thalassemia, Duhok, XmnI polymorphism, Iraq

β -thalassemia (thal) is an autosomal recessive genetic disease, which is among the most frequently encountered inherited hematological disorders in Iraqis,

including those from Duhok province in the extreme north.¹⁻⁴ β -thalassemia may present as a severe transfusion dependent thalassemia major phenotype (with

* Professor and Head of Department of Pathology, Faculty of Medical Sciences, School of Medicine, University of Duhok.

** Lecturer of Molecular Biology, Scientific Research Center, Faculty of Science, University of Duhok.

*** Demonstrator, Scientific Research Center, Faculty of Science, University of Duhok.

**** Registrar, Department of Hematology, Azadi Teaching Hospital, Duhok, Iraq.

***** Lecturer and Head of Department of Hematology, Central Laboratory, College of Medicine, University of Sulaymaniyah, Iraq.

***** Director, Thalassemia Care Center, Duhok, Iraq.

Correspondence author: Prof Nasir Al-Allawi, School of Medicine, faculty of medical Sciences, University of Duhok, Iraq. Tel: +964750455 1494. Email : nallawi@gmail.com

homozygous or compound heterozygous genotypes), or a relatively asymptomatic thalassemia minor (with heterozygous genotype) or an intermedia phenotype with intermediate clinical severity and a variable genotype.⁵ Several genetic factors have been implicated in modulating the severity of symptomatic β -thalassemia, including the type of β -gene mutations, the concomitant α -gene status and increased γ -gene production.⁶ Among the most important genetic factors involved in the latter, is a mutation causing a C to T base pair substitution at the -158 position in promoter region of the γ -globin gene. This mutation leads to the creation of a digestion site for the restriction enzyme *Xmn* I (*Xmn* I polymorphism).⁷ Although *Xmn* I polymorphism contribution to the phenotypic variation in β -thalassemias has been investigated and documented in several populations, no such study was reported among Iraqis. The aim of the current study is to address the latter issue through studying a group of β -thalassemia major and intermedia patients.

METHODS

A total of 107 cases of symptomatic β -thalassemia patients were recruited from the thalassemia care center in Duhok. They included 55 patients with thal major (TM) and 52 with thal intermedia (TI). A careful review of the clinical and laboratory records of all enrolled patients was undertaken. The diagnoses of thal major or intermedia were based on the results of initial investigations at diagnosis as well as clinical follow up and transfusion dependence.

A sample of 7.5 mL of venous blood was taken and distributed between 3 EDTA tubes. One sample was used to perform a full blood count via an electronic hematology analyzer (Beckman coulter – USA), another to perform high performance liquid chromatography (HPLC) (VARIANTTM; Bio-Rad Laboratories, Hercules, CA, USA) for

quantitation of Hb A, A2 and F, while the third sample tube was stored at -20°C and was thereafter used for DNA extraction using a phenol-Chloroform method.⁸

The extracted DNA was then amplified for a 650 bp sequence in the promoter region of the γ -globin gene. This amplification was performed using an AB 2720 thermocycler (Applied Biosystems-USA) applying the following primers: Forward 5' AAC TGT TGC TTT ATA GGA TTT T3' and Reverse 5' AGG AGC TTA TTG ATA ACT CAG AC 3'. The PCR program used started with an initial denaturation for 2 min at 94°C, followed by 30 cycles of denaturation at 95°C for 1 min, annealing 60°C for 1 min, and extension 72°C for 1.5 min. Thereafter, a final extension for 5 minutes at 72°C.⁸

The 650 bp amplicon was then digested with the enzyme *Xmn* I according to the manufacturer's instructions (Promega-USA), and the digestion products were run on a 2% agarose gel and visualized using ultraviolet transilluminator after staining with ethidium bromide.

This study was approved by the ethical committee at the Scientific Research Center, University of Duhok.

Statistical analysis utilized the SPSS software program. Chi square (with Yates correction) and Mann Whitney U test were used when applicable. $P < 0.05$ was considered significant.

RESULTS

The results of molecular studies showed that out of the 52 patients with thal intermedia: 10 patients (19.2%) were found to be homozygous for *Xmn* I (+/+), compared to only 1/55 in the thal major group (1.8%); while 17/52 (32.7%) of thal intermedia and 12/55 (21.8%) of thal major were heterozygous for it (+/-). Overall, those who were carriers of the *Xmn* I polymorphism (whether +/+ and +/-) and particularly those homozygous for it

(+/+) were significantly more frequent among thal intermedia group ($p=0.0048$ and 0.0028 respectively). Figure 1 shows the RCR-RFLP results.

The results also revealed that out of the total 104 chromosomes in the thal intermedia group 37 (35.6%) had the *Xmn* I polymorphism, compared to 14 (12.7%) of the 110 thal major chromosomes. A finding which was highly significant ($p=0.0002$).

Median transfusion requirements/year were 0, 3.5 and 11, in those with *Xmn* I (+/+), (+/-) and (-/-) categories respectively regardless whether they were in thal major or intermedia categories (with significantly lower requirements in (+/+) and (+/-) when compared to (-/-) with p of 0.004 and 0.020 respectively).

When median hemoglobin concentration (Hb), Hb F and Hb A2 were compared between those with the *Xmn* I polymorphism and those without it in the 50 thal intermedia patients who did not receive any transfusions in the past 2 months, it was found that there were significant higher Hbs in those with (+/+) when compared with the (-/-) ($p=0.044$) but no significant differences between (+/-) and the (-/-) genotypes ($p=0.072$). Furthermore, Hb F was significantly higher in those with (+/+) and (+/-) when compared to those with (-/-) genotype ($p=0.029$ and 0.022 respectively). Hb A2 on the other hand, was significantly lower in those with (+/+) and (+/-) compared to those without the polymorphism ($p=0.006$ and 0.009 respectively). Table 1 outlines the median Hb, HbF and Hb A2 in association with *Xmn* I polymorphism.

Table 1. Median Hb, Hb A2 and F levels in relevance to the *Xmn* I polymorphism in 50 Thal Intermedia patients from Duhok

Parameter	<i>Xmn</i> I status (No.)		
	+/+	+/-	-/-
	9	17	24
Hb (g/dl)	9.9	8.2	8.6
Hb A2 (%)	3.5	3.8	5.95
Hb F (%)	91.7	45.5	16.25



Figure 1. Represent 2% gel electrophoresis for PCR-RFLP analysis of *Xmn* I polymorphism. Lane 1: 100 bp DNA ladder; Lane 2: 650 bp fragment from a patient with *Xmn* I (-/-) genotype; Lanes 3: 650 bp, 450 bp and 200 bp fragments from patients heterozygous for the *Xmn* I (+/-) genotype; Lanes 4: 450 bp and 200 bp digested fragments from a patient with *Xmn* I (+/+) genotype; Lane 5: Undigested 650 bp amplified sequence

DISCUSSION

Several factors have been implicated in modulating the severity of β -thalassemia, including those which lead to increase in γ -globin chain production. *Xmn* I polymorphism has been reported to be associated with 3-11 folds increase in G_γ -globin chain production, by increasing the rate of the transcription of the gene, in conditions characterized by hemopoietic stress.^{7,9,10} The subsequent increase in cellular Hb F content offers a selective survival advantage to the cells and thus ameliorating the disease.^{7,9} Its contribution to the molecular basis of thalassemia intermedia varies in different populations, with the highest rates reported among Iranian and the least among the Chinese.^{11,12}

The findings of the current study show that among Kurdish thalassemic patients, *Xmn* I polymorphism is more frequently encountered in those with thal intermedia when compared to those with thal major, and that homozygosity (+/+) was significantly higher among TI. This

indicates that *Xmn* I polymorphism is an important modulator of thal severity among the Iraqi Kurds. This is strongly supported by the findings that the median transfusions per year decreased from a median of 11/year in those with *Xmn* I (-/-), to 3.5/year in the *Xmn* I (+/-) to zero/year for the (+/+) category, an observation which was found to be significant. However, in patients with thal intermedia, hemoglobin was only significantly higher in those with *Xmn* I (+/+), but not in those with (+/-) genotypes, versus the (-/-) genotype. Previous investigators have suggested that significant amelioration of thalassemia was more likely in (+/+) than (+/-) genotypes.⁶

When compared to other studies on thalassemia intermedia, our 19.2% *Xmn* I (+/+) figure is near to those reported from Lebanon at 21.9% by Qatanani and coworkers (2000), Mediterraneans and Asian-Indians by Ho and coworkers (1998) and within the range given for Indians of 12.5-27.4%, but was much less than the high figure of 40% reported by Neishabury and colleagues (2008) among Iranian TI patients.¹¹⁻¹⁶ In all the above mentioned populations it appears that *Xmn* I (+/+) polymorphism is an important contributor to the molecular basis of TI, which is in contrast to almost absent role of this polymorphism among the Chinese TI, where (+/+) polymorphism is seen in <1% of TI patients.¹²

Hemoglobin, Hb F and A2 could only be compared in thal intermedia patients (50/52) who did not receive a recent transfusion. Interestingly and to further support the role of *Xmn* I polymorphism as a modulator of Hb F, it was shown that Hb F was significantly higher in those with the polymorphism whether in homo or heterozygous state than non-carriers. However, the remarkable variation in Hb F levels in those with *Xmn* I polymorphism and those without it, indicates that those with (-/-) are more likely to have another major ameliorating factor which is likely

to be the inheritance of mild β -thal mutations (homozygous or compound heterozygous) or heterozygosity to such mutations. Both latter categories are associated with lower Hb F levels.⁵

In conclusion, this study has shown that *Xmn* I polymorphism is an important β -thal genetic modulator in Iraqi Kurdish thal patients, and that this polymorphism most likely exerts its effect through increasing Hb F. However, further studies on other genetic modulators of the phenotype of symptomatic β -thal are warranted. Among those requiring further scrutiny are types of the β -thal mutations and concomitant α -thal.^{5,9,14}

REFERENCES

1. Yahya HI, Khalel KJ, Al-Allawi NAS, Hilmi F. Thalassaemia genes in Baghdad, Iraq. East Mediterr Health J. 1996 ; 2 (2); 315-9.
2. Hassan MK, Taha JY, Al-Noama LM, Widad NM, Jasim SN. Frequency of hemoglobinopathies and Glucose-6-Phosphate dehydrogenase deficiency in Basra. East Mediterr Health J. 2003;9 (1-2);45-54.
3. Jalal S, Al-Allawi N, Faraj A, Ahmed N. Prevalence of hemoglobinopathies in Sulymani – Iraq. Duhok Med J. 2008;2(1):71–9.
4. Al-Allawi NA, Al-Dousky AA. Frequency of haemoglobinopathies at premarital health screening in Dohuk, Iraq: implications for a regional prevention programme. East Mediterr Health J. 2010; 16(4): 381-5.
5. Weatherall DJ, Clegg JB. The thalassaemia syndromes. 4th ed. Oxford: Blackwell scientific Publications; 2001.
6. Winichagoon P, Fucharoen S, Chen P, Wasi P. Genetic factors affecting clinical severity in β -thalassemia syndromes. J Pediatr Hematol Oncol. 2000; 22(6): 573-80.
7. Gilman JG, Huisman THJ. A sequence variation associated with elevated fetal

- ^Gγ-globin production. Blood. 1985; 66: 783-7.
8. Old J, Traeger-Synodinos J, Galanello R, Petrou M, Angastiniotis M. Prevention of thalassaemia and other hemoglobin disorders. Vol 2. Nicosia: TIF publications; 2005.
 9. Thein SL. Genetic modifiers of the β-haemoglobinopathies. Brit J Haematol. 2008; 141: 357-366.
 10. Sampietro M, Thein SL, Contreras M, Laszlo P. Variation of HbF and F-cell number with the G-gamma *Xmn* I (C-T) polymorphism in normal individuals. Blood. 1992; 79(3): 832-3.
 11. Neishabury M, Azarkeivan A, Oberkanins C, Esteghamat F, Amirizadeh N, Najmabadi H. Molecular mechanisms underlying thalassemia intermedia in Iran. Genet Test. 2008; 12(4): 549-56.
 12. Chen W, Zhang X, Shang X, Cai R, Li L, Zhou T, et al. The molecular basis of beta-thalassemia intermedia in Southern China: genotypic heterogeneity and phenotypic diversity. BMC Med Genet. 2010, 11:31.
 13. Qatanani M, Taher A, Koussa S, Naaman R, Fisher C, Rugless M, et al. β-thalassaemia intermedia in Lebanon. Eur J Haematol. 2000; 54: 237-44.
 14. Ho PJ, Hall GW, Luo LY, Weatherall DJ, Thein SL. Beta-thalassaemia intermedia: is it possible consistently to predict phenotype from genotype. Br J Haematol. 1998; 100(1): 70-8.
 15. Panigrahi I, Agarwal S, Pradhan M, Choudhry DR, Choudhry VP, Saxena R. Molecular characterization of thalassemia intermedia in Indians. Haematologica. 2006; 91(9): 1279-80.
 16. Nadkarni A, Gorakshakar AC, Lu CY, Kirshnamoorthy R, Ghosh K, Colah R, et al. Molecular pathogenesis and clinical variability of β-thalassaemia syndromes among Indians. Am J Haematol. 2001; 68(2):75-80.

پوخته

مشه شیوهییا Xmn I ل نه خوشین دهریایی سپی ناوه راست ل جورئ بیتا ل ده فراه دهوکی

پیشگی و نارمانج: گه له گهورکه ریڼ زکماکی یڼ هاتینه نیشانکرڼ بو توشبوونی ب جورهیڼ شیوهی یڼ نه خوشیا تالاسیمیا بیتا یا ب نیشان. ژ یڼ مشه هاتینه نیشانکرڼ ل روژه لاتا نافه راست مشه شیوهییا Xmn I. چ فه کولینا بابه تی گریدانا فی مشه شیوهیڼ دگه ل جورهیڼ شیوهی یڼ نه خوشین تالاسیمیا بیتا یڼ عراقی نه وه رگرتیه.

ریکڼ فه کولینی: سه رجه می 107 نه خوشین تالاسیمیا بیتا هاتنه وه رگرتن دفی فه کولینی دا (52 ژ جورئ تالاسیمیا نافین و 55 یڼ تالاسیمیا مه زن). پیداجوونه ک د تومارین وان دا هاته کرن و تاقیکرنا ته مام یا خوینی و هیموگلوبینی A و A2 و F بو هاته کرن بریكا HPLC و DNA هاته فافارتن. نه ف DNA هاته بکارئینان بو ده ستنیشانکرنا مشه شیوهییا Xmn I بریكا PCR-RFLP.

نه دجام: هاته دیتن کو Xmn I یی وه هه ف (+/+) هه بو ل ده ف 19.2٪ و 1.8٪ ژنه خوشین تالاسیمیا نافین و تالاسیمیا مه زن لدویف ئیک (p= 0.0028). حاله تی Xmn I یی نه وه هه ف (-/+) هاته دیتن ل ده ف 32.7 و 21.8٪ لدویف ئیک. بلندترین ئاستی هیموگلوبینی F هاته دیتن ل ده ف نه وین (+/+) و کیمترین هاته دیتن ل ده ف نه وین (-/-) و نه ف پیزانینه یا گرنه بو (p= 0.029).

ده رنه دجام: مشه شیوهییا Xmn I گهورکه ره کی زکماکی یی گرنه بو دژواریا تالاسیمیا بیتا ل ده ف کوردین عراقی و گریدان یا هه ی دگه ل بلندبوونا هیموگلوبینی F. پتر فه کولین لسه ر گهورکه ریڼ زکماکی پیدفی نه بو ده ستنیشانکرنا به شداربوونا وان د جورهیڼن شیوهی یڼ فی نه خوشیا زکماکی یا گرنه.

الخلاصة

تعدد أشكال Xmn I في مرضى بيتا تالاسيميا في دهوك

خلفية واهداف البحث: يوجد العديد من المعدلات الوراثية المؤثرة في الاشكال المظهرية لمتلازمة فقر دم البحر المتوسط ومن المعدلات المهمة في شرقي البحر المتوسط هو polymorphism XmnI و لكن لا توجد دراسة بخصوص المساهمة النسبية لهذا المعدل للمرضى العراقيين.

طرق البحث: تم دراسة مجموعه من 107 مرضى فقر دم البحر الابيض المتوسط (52 ثالاسيميا وسطية و 55 ثالاسيميا كبرى) . وقد تم مراجعة ملفاتهم المرضية ثم عمل صورة دم كاملة و تحديد نسب A2 و F بطريقة HPLC و استخلاص الدنا . ومن ثم استعمل الدنا لتحديد وجود المعدل XmnI بطريقة PCR-RFLP.

النتائج: وجد ان المعدل XmnI متمائل الزيجة (+/+) وجد في 19.2 % و 1.8 % من مرضى الثالاسيميا الوسطية والكبرى على التوالي ($p=0.0028$). أما متباين الزيجة XmnI (-/+) فقد وجدت في 32.7% و 21.8 % على التوالي. وقد وجد انه ومن بين 107 مريضا مشمولوا بالدراسة فإن الأنماط الوراثية (+/+), (-/+), و (-/-) بالتتابع إرتبطت بالأحتياج لنقل دم سنوي وسيط 0, 3.5 و 11 على التوالي مع احتياج اقل للمرضى الحاملين للأنماط +/+ و +/- مقارنة مع غير الحاملين (0.02 و $p=0.004$) على التوالي. وقد وجد ان اعلى نسبة Hb F كانت موجودة في الاشخاص حاملين النمط (+/+) وأقل نسبة في الاشخاص حاملين نمط (-/-) و بمستوى معنوي مهم ($p=0.029$).

الاستنتاجات: أن النمط الوراثي I_A Xmn هو معدل وراثي مهم لتحديد شدة فقر البحر الابيض المتوسط بين الأفراد العراقيين، و ان وجوده مرتبط بارتفاع مستوى Hb F عند المقارنة مع غير الحاملين له . ويبدو ان هناك معدلات وراثية أخرى تحتاج للدراسة لتحديد دورها في المساهمة في كشف الاختلافات المظهرية في هذا المرضى الوراثي المهم.

INCIDENCE AND ANTIBIOTIC SUSCEPTIBILITY PATTERNS OF
PSEUDOMONAS AERUGINOSA IN BURNS INFECTIONS IN DUHOK CITY

NAJIM A. YASSIN, PhD*
JANAN M. SALIH, MSc**
BLIND H. ABDULLA, MSc***

Submitted 4 May 2011; accepted 26 Feb 2012

ABSTRACT

Background and objectives *Pseudomonas aeruginosa* is an opportunistic pathogen causing severe, acute and chronic nosocomial infections. The organism is generally resistant to numerous antimicrobial agents, making the treatment of infections further difficult. The aim of this study was to determine the incidence of *Pseudomonas aeruginosa* involved in burns infections. Also, the susceptibility and resistotyping patterns of the isolates to commonly prescribed antibiotics were studied.

Methods During a period of six months between July and December, 2010, a total of 159 samples from burns infections, using sterile cotton swabs, were collected from Burns Hospital in Duhok city. The samples were plated on Blood agar and MacConkey agar and the isolates were identified by routine procedures. Antibiotics susceptibility and resistant profiles to 14 commonly prescribed antibiotics were performed by the disc diffusion method using Mueller-Hinton agar.

Results Out of the 159 samples collected from burns infections, 116 samples were showed bacterial growth, 76 (47.7%) were *Pseudomonas aeruginosa*, followed by *Klebsiella pneumoniae* 20 (12.5%), *Escherichia coli* 10 (6.2%), *Staphylococcus aureus* 8 (5 %), *Staphylococcus epidermidis* 2 (1.2%), and no growth 43 (27 %). The results showed that the occurrence of *Pseudomonas aeruginosa* was higher than the other groups of bacteria. The sensitivity pattern of *Pseudomonas aeruginosa* revealed that the organism was highly sensitive to imipenem (98.6%) followed by piperacillin (60.5), ciprofloxacin (57.8), and amikacin (48.6%). On other hand, chloramphenicol (19.7%), doxycycline (10.8%), ceftazidime (10.8%), erythromycin (6.5%), gentamicin (3.9%), cefotaxime (3.9%), amoxiclav (3.9%), tetracycline (3.9%), vancomycin (3.9%) and cefixime (2.6%) showing the lowest percentages sensitivity. Resistant profiles were determined. A total of 12 different resistotype patterns were obtained; common resistotype were 5 and 11.

Conclusions This study shows that there is an increased rate of incidence of *Pseudomonas aeruginosa* in burns infections and most of these isolates were multi-drug resistant and showed different resistotyping patterns.

Duhok Med J 2012;6(1): 8-16.

Key words: *Pseudomonas aeruginosa*, Multidrug-resistance, Burns infection, Resistotyping patterns, Antibiotics

Pseudomonas aeruginosa is a gram-negative rod measuring 0.5 - 0.8 µm by 1.5 to 3.0 µm, commonly found in soil and water.¹ *Pseudomonas aeruginosa* can

* Lecturer of Medical-Microbiology, School of Medicine, Faculty of Medical Sciences, Duhok University, Kurdistan Region, Iraq.

** Assistant Lecturer of Medical-Microbiology, School of Medicine, Faculty of Medical Sciences, Duhok University, Kurdistan Region, Iraq.

*** Assistant Lecturer of Medical-Microbiology, School of Nursing, Faculty of Medical Sciences, Duhok University, Kurdistan Region, Iraq.

Correspondence author: Najim A. Yassin, School of Medicine, Faculty of Medical Sciences, Duhok University, Kurdistan Region, Iraq. E.mail: najim56@yahoo.com

cause infections virtually anywhere in the body, but urinary tract infections, pneumonia (especially in cyst fibrosis patients) and wound infections (especially burns) predominate. From these sites, the organism can enter the blood, causing sepsis. *Pseudomonas aeruginosa* have a remarkable ability to withstand disinfectants, this accounts in part for their role in hospital-acquired infections, they have been found growing in hexachlorophene-containing soap solutions, in antiseptic and in detergents.² Almost all the clinical cases of *Pseudomonas aeruginosa* infection can be associated with the compromise of host defense such as burn patients. While many cases of *Pseudomonas aeruginosa* infection can be attributed to general immuno-suppression e.g. AIDS patients.^{3,4}

Most pseudomonad burns infections are established through colonization of the burns wound by the patient's own flora or from the environment. Most fatalities (usually arising from septicemia) are associated with full-thickness burns, and there is a strong correlation with the percentage area of the burn.⁵ Patients with burns infected with *Pseudomonas aeruginosa* have an increased mortality rate and longer hospital care compared to non-infected patients. They also have an increased number of surgical procedures and higher associated antibiotic costs.⁶ Burn hospitals often harbor multidrug-resistant *Pseudomonas aeruginosa* that can serve as the source of infection. *Pseudomonas aeruginosa* has been found to contaminate the floors, bed rails, and sinks of hospitals, and has also been cultured from the hands of nurses.⁷

Concerning multi-drug resistance, Hsueh et al 1998, reported multi-drug-resistant strain of *Pseudomonas aeruginosa* over a period of several years was carried by some patients asymptotically through several rounds of antibiotic treatment for *Pseudomonas aeruginosa* infections.⁸ This scenario can be worse during the spread of

Pseudomonas aeruginosa from one patient to another; the persistence of this strain takes place in patients throughout several courses of antibiotic treatment.⁵ It has been proved that during admission of patients in burn centers, a limited number of common strains cross-contaminate burn victims mostly when their lesions scrubbed in the bathroom.⁹

Pseudomonas aeruginosa exhibits resistance to a variety of antimicrobials including beta lactams. Carbapenems are often used as antibiotics for treatment of infections caused by beta lactam resistant *Pseudomonas aeruginosa*.¹⁰ The aim of the present study was to find out the incidence and antibiotic susceptibility of *Pseudomonas aeruginosa* isolates recovered from the burns infections, also establishment the resistotype patterns of these isolates.

METHODS

The specimens were collected from patients aseptically with sterile cotton wool swab suffering from burns infections at burns hospital in Duhok city from July and December 2010. Swabs being routinely processed by the Department of Laboratory Service at Burns Hospital. Several media and tests were used for the isolation; identification and testing the susceptibility of the isolates for commonly used antibiotics. The media used are:

Blood agar (with 5-7% defibrinized blood), MacConkey agar, chocolate agar, nutrient agar, Mannitol salt agar. Simmons citrate agar, kligler Iron Agar (KIA), Mueller-Hinton agar, Sulfide formation indole production, motility Test (SIM), Nutrient agar, Methyl Red-Voges Proskauer broth, Thioglycollate broth, Coagulase, Catalase, Urease, Oxidase tests were used for the identification. All of the above media and reagents were obtained from (Difco. USA).

The media were prepared according to manufacturers instructions in 500 mL bottle and sterilized by autoclaving at

121°C for 20 minutes. All wound swabs collected for bacteriological investigations during the period of this study were treated according to established method of treating wound swabs.¹¹ Gram stain preparations were made from all the swabs the plates were incubated at 37°C for 18-24 hours in an incubator. The plates were read the following day but extended to 48 hours if there was no bacterial growth within 24 hours. Isolated colonies were subjected to Gram staining technique and biochemical tests for identification.

Antibiotic sensitivity tests were carried out on isolated and identified colonies of *Pseudomonas aeruginosa* using commercially prepared antibiotic sensitivity disc using Kirby-Bauer method.¹²

RESULTS

A total of 159 samples were collected from burns hospital, all specimens were directly transferred to the microbiology laboratory and cultured to the appropriate media (as described in methods).

Table 1 shows the most causative agents of burns infections were *Pseudomonas aeruginosa* 76 isolates (47.7%), followed by *Klebsiella pneumoniae* 20 isolates (12.5%). The lowest causative agents of burns infections

were *Escherichia coli*, *Staphylococcus aureus*, and *Staphylococcus epidermidis*.

Table 1. Frequency and percentage of microorganisms isolated from patients in burns infections

Microorganism	Number of isolation (%)
<i>Pseudomonas aeruginosa</i>	76 (47.7)
<i>Klebsiella pneumoniae</i>	20 (12.5)
<i>Escherichia coli</i>	10 (6.2)
<i>Staphylococcus aureus</i>	8 (5.0)
<i>Staphylococcus epidermidis</i>	2 (1.2)
No growth	43 (27.0)
Total	159 (100)

Table 2 shows sensitivity patterns of *Pseudomonas aeruginosa* isolated from patients with burns infections. The organism was sensitive to imipenem followed by piperacillin, ciprofloxacin, and amikacin.

Table 3 shows the resistotyping patterns of *Pseudomonas aeruginosa* isolates to commonly used antibiotics. It was found that all of the isolates were multiple resistant, i.e. resistant to more than one antibiotic. The obtained results of resistotyping patterns (resistant profiles) revealed that 58 isolates were multiple drug resistant to antibiotics and 12 different patterns were found.

Table 2. Susceptibility test of *Pseudomonas aeruginosa* isolated from patients in burns infections

Antibiotics	Symbol	Disc potency (µg)	Susceptibility (%)
Imipenem	Imp	10	98.6
Piperacillin	PRL	100	60.5
Ciprofloxacin	CIP	5	57.8
Amikacin	AK	30	48.6
Chloramphenicol	C	10	19.7
Ceftazidime	CAZ	30	10.8
Doxycycline	DOX	30	10.8
Erythromycin	E	15	6.5
Gentamicin	CN	10	3.9
Cefotaxime	CTX	30	3.9
Vancomycin	VA	30	3.9
Amoxi-clav	AMV	5	3.9
Tetracycline	TE	30	3.9
Cefixime	CFM	30	2.6

Table 3. Resistotyping patterns of *Pseudomonas aeruginosa* isolates

Resistotyping patterns	Resistance Spectrum Phenotype	No. of isolates (%)
Resistotype 1	DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC	2 (3.44)
Resistotype 2	AK, CTX, CAZ, TE, CFM, VA, E, CN, AMC	3(5.1)
Resistotype 3	CIP, DOX, CTX, TE, CFM, VA, E, CN, AMC, C	4(6.8)
Resistotype 4	AK,PRL, CTX, CAZ, TE, CFM, VA, E, CN, AMC	2(3.4)
Resistotype 5	DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC,C	9(15.9)
Resistotype 6	PRL, DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC	4(6.9)
Resistotype 7	AK, DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC	3(5.1)
Resistotype 8	CIP, DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC, C	5(8.2)
Resistotype 9	AK, CIP, DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC	7(12)
Resistotype 10	PRL, CIP, DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC	5(8.2)
Resistotype 11	AK, PRL, DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC	9(15.9)
Resistotype 12	AK, PRL, CIP,DOX, CTX, CAZ, TE, CFM, VA, E, CN, AMC	5(8.2)
Total		58(100)

DISCUSSION

The burn wound is considered one of major health problem in the world, and infection is one of the frequent and severe complications in patients who sustained burns.^{13,14} The burn wound represents a susceptible site of opportunistic colonization by organisms of endogenous and exogenous origin. Patient factors such as age, extent of injury and depth of burns in combination with microbial factors determine the likelihood of invasive burn wound infection.¹⁵

In our study *Pseudomonas aeruginosa* was found in burns infections 76 (47.7%), be the most common organism isolated during 6 months from burns hospital, followed by *Klebsiella pneumoniae* (12.5.6%). This is in agreement to other studies which showed *Pseudomonas* as the most common infective organism in burns patients. Naser et al 2003, from Cairo, Egypt has reported *Pseudomonas aeruginosa* was the most frequent isolate (21.6%), followed by *Klebsiella pneumoniae* (15.2%) and *Staphylococcus* 11.6%.¹⁶ Taneja et al 2004, from Chandigarh, India have reported *Pseudomonas aeruginosa* as the most frequent isolates (54.2%) followed by *Staphylococcus aureus* (20.8%). Other study in india indicated that the *Pseudomonas aeruginosa* was predominant in burns infections.¹⁷

Nowadays, the prevalence of *Pseudomonas aeruginosa* and the new resistant strains continue in both community-acquired pathogens and hospital originated infections.¹⁸ In our study most of *Pseudomonas aeruginosa* isolates were highly sensitive to imipenem (98.6%) followed by piperacillin (60.5%), ciprofloxacin (57.8%), and amikacin (48.6%). In a study conducted by Revathi et al, *Pseudomonas aeruginosa* was most susceptible to ceftizidime (83%) and cefoperazone (82%).¹⁹ In a study carried out in Turkey by Inan et al, isolated 68% of *Pseudomonas aeruginosa* strains and 60-83% of the antibiotics resistant strains were from ICU patients. In the same study, resistance was detected against ceftazidime 34%, imipenem 26%, gentamicin 67%, and amikacin 26%.²⁰

Our study showed that the susceptibility rate of *Pseudomonas aeruginosa* to gentamicin was very low as 3.9%. Reports of the susceptibility of *Pseudomonas aeruginosa* to gentamicin have ranged from as low as 49.8% in Greece, to 70% in Turkey, to as high as 96.6%, in the United Kingdom.²¹ In Trinidad and Tobago, 80 and 78.4% of isolates were susceptible to ceftazidime and gentamicin respectively.²² Report from France have shown *Pseudomonas aeruginosa* susceptibility rates of 78.5 and 61.7% to ceftazidime and ciprofloxacin, respectively.²³ Increased resistance was

observed in Russia where only 25% of isolates were susceptible to gentamicin.²⁴ While in Bangladesh 49 and 79% of isolates were susceptible to tobramycin and ciprofloxacin respectively.²⁵

Consistent with these findings, resistance to gentamicin of *Pseudomonas aeruginosa* is increasing progressively in our country. The most important risk factors are obvious, such as excessive consumption of antibiotics exerting selective pressure on bacteria, the frequent use of invasive devices and relative density of a susceptible patient population in burns units.²⁶

Ciprofloxacin resistance rate was 42.2% in our study, 27.4% in Turkey,²⁰ 32% in Spain,²⁷ 31.9% in Italy,²⁸ 26.8% in Latin America,²⁹ 31% in India,¹⁷ and 100% in Iran.³⁰ Thus, in burns infections, empirical antibiotic treatments should be avoided and treatment should be carried out using antibiotic susceptibility tests. While piperacillin resistance rate was 39.5% in our findings, 10% in Spain,²⁷ 12% in Italy,²⁸ 14% in Latin America,²⁹ 28.7% in Turkey,²⁰ 20% in India,¹⁷ and 100% in Iran.³⁰ Most studies conducted in the world shows that most common drugs are resistant to the organism isolated like ampicillin, erythromycin and cefotaxime.^{31,32} These findings are consistent with our findings.

Resistotyping is a phenotypic method that consists of testing bacterial strains against a set of arbitrarily chosen antibiotics, whereby, a resistance pattern that is characteristic of a strain is generated and, is believed to describe the isolates for epidemiological purposes. Obtained results of this study revealed that the 58 isolates of *Pseudomonas aeruginosa* from burns infections were belonged to 12 distinct resistotype patterns. Resistotype 5 and 11 have much higher frequency rate comprising 15.9% (for each one) of the isolates. Studies in Spanish,³³ Brazil,³⁴ Iran,³⁵ and Iraq,³⁶ conducted resistant profiles of strains *Pseudomonas aeruginosa*. Moreover, those isolates, from

various patients, were identical on the basis of disk susceptibility patterns, indicating relatedness among them. In general, this simple typing system provide discriminatory between strains and able to determine relatedness among isolates of *Pseudomonas aeruginosa* in order to tracing the source of infections in our environment.

This study concluded that there is an increased rate of incidence of *Pseudomonas aeruginosa* in burns wounds infections followed by *Klebsiella pneumoniae*, *Escherichia coli*, *Staphylococcus aureus*, and *Staphylococcus epidermidis*. This is in agreement with survey studies carried out in various hospitals. The infection appears to be common in burns hospitals owing to the burns units are a very susceptible habitat for bacterial colonization and excessive use of empiric antibiotic treatment. The reason for this increase in burns infections rate with prolonged hospitalization is primarily due to colonization of patients with hospital-acquired resistant microorganisms. Resistance of *Pseudomonas aeruginosa* to antibiotics is very definitely associated with overuse of broad-spectrum antibiotics in hospitals, lead to different resistotyping patterns of antibiotics have been determined. Therefore, new and more effective antibiotics may be needed.

REFERENCES

1. Krieg N. Bergey's Manual of systematic bacteriology. 4th ed. Baltimore (MD): Williams & Wilkins; 1984.
2. Warren L. Review of medical microbiology and immunology. 10th ed. New York: McGraw-Hill; 2008.
3. Franzetti F, Cernuschi M, Esposito R, Moroni M. *Pseudomonas* infections in patients with AIDS and AIDS-related complex. J Intern Med. 1992;231(4):437-43.
4. Kielhofner M, Atmar RL, Hamill RJ,

- Musher DM. Life-threatening *Pseudomonas aeruginosa* infections in patients with human immunodeficiency virus infection. Clin Infect Dis. 1992;14(2):403-11.
5. Smith RP. Skin and soft tissue infections due to *Pseudomonas aeruginosa*. In: Baltch AL, Smith RP, editors. *Pseudomonas aeruginosa* infections and treatment. New York: Marcel Dekker; 1994. p. 327-69.
 6. Tredget EE, Shankowsky HA, Rennie R, Burrell RE, Logsetty S. *Pseudomonas* infections in the thermally injured patient. Burns. 2004;30(1):3-26.
 7. Chitkara YK, Feierabend TC. Endogenous and exogenous infection with *Pseudomonas aeruginosa* in a burns unit. Int Surg. 1981;66(3):237-40.
 8. Hsueh PR, Teng LJ, Yang PC, Chen YC, Ho SW, Luh KT. Persistence of a multidrug-resistant *Pseudomonas aeruginosa* clone in an intensive care burn unit. J Clin Microbiol. 1998;36(5):1347-51.
 9. Japoni A, Farshad S, Alborzi A, Kalani M, Mohamadzadegan R. Comparison of arbitrarily primed-polymerase chain reaction and plasmid profiles typing of *Pseudomonas aeruginosa* strains from burn patients and hospital environment. Saudi Med J. 2007;28:899- 903.
 10. Lee K, Lim YS, Yong D, Yum JH, Chong Y. Evaluation of the Hedge test and the imipenem-EDTA double-disk synergy test for differentiating metallo beta lactamase-producing isolates of *Pseudomonas spp.* and *Acinetobacter spp.* J Clin Microbiol. 2003;41(10):4623-9.
 11. Masaadeh H A, Jaran A S. Incident of *Pseudomonas aeruginosa* in post-operative wound infection. Am J Infect Dis. 2009;5(1):1-6.
 12. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. Am J Clin Pathol. 1966; 45(4):493-6.
 13. Zorgani A, Zaidi M, Ranka R, Shaher A. The pattern and outcome of septicemia in burns intensive care unit. Ann Burns Fire Disasters. 2002; 15(4):179-81.
 14. Steinsträsser L, Thies AH, Rabstein S, Steinau HU. Typical bacteria in an intensive care burn unit in severely burned patients and their importance with regard to mortality: retrospective study 1995-2004. Handchir Mikrochir Plast Chir. 2007;39(5):338-44. [Article in German]
 15. Imran M, Faheem M, Aslam V, Hakeem A, Inayat U, and shah A. wound infection and culture sensitivity pattern in pediatric burn patients. JMPI. 2009;23(4):304-8.
 16. Naseer S, Mabrouk A, Maher A. Colonization of burn wounds in Ain shams University Burn Unit. Burns. 2003; 29(3):229-33.
 17. Taneja N, Emmanuel R, Chari PS, Sharma M. A prospective study of hospital-acquired infections in burn patients at a tertiary care referral centre in North India. Burns. 2004;30(7):665-9.
 18. Maniatis AN, Trougakos IP, Katsanis G, Palermos J, Maniatis NA, Legakis N. Changing patterns of bacterial nosocomial infections: a nine-year study in a general hospital. Chemotherapy. 1997; 43(1):69-76.
 19. Revathi G, Puri J, Jain BK. Bacteriology of burns. Burns. 1998; 24(4):347-9.
 20. İnan D, Ögünç D, Günseren F, Çolak D, Mamikoğlu L, Gültekin M. The resistance of *Pseudomonas aeruginosa* strains isolated from nosocomial infections against various antibiotics. Mikrobiyol Bult. 2000;34(3-4): 255-60.
 21. Van Landuyt HW, Boelaert J, Glibert B, Gordts B, Verbruggen AM. Surveillance of aminoglycoside resistance. European data. Am J Med.

- 1986;80(6B):76-81.
22. Fitzroy A, Orrett MD. Antimicrobial susceptibility survey of *Pseudomonas aeruginosa* strains isolated from clinical sources. *J Nat Med Assoc.* 2004;96(8):1065-9.
23. Bertrand X, Thouverej M, Patry C, Balvay P, Talon D. *Pseudomonas aeruginosa*: antibiotic susceptibility and genotypic characterization of strains isolated in the intensive care unit. *Clin Microbiol Infect.* 2001;7(12):706-8.
24. Stratcheunski LS, Russian NPRS Group, Kozlov RS, Rechedko GK, Stetsiouk OU, Chavrikova EP. Antimicrobial resistance patterns among gram -negative bacilli isolated from patients in intensive care units: Results multicentre study in Russia. *Clin Microbiol Infect.* 1998;4(9):497-507.
25. Ansary SP, Haque R, Faisal AA, Chowdhury MS. Resistance pattern of *Pseudomonas aeruginosa* occurring in northern part of Bangladesh. *Trop Doct.* 1994;24(4):188.
26. Savaş L, Duran N, Savaş N, Önlen Y, Ocak S. The prevalence and resistance patterns of *Pseudomonas aeruginosa* in intensive care units in a University Hospital. *Turk J Med Sci.* 2005;35:317-22.
27. Bouza E, Garcia-Gorrote F, Cercenado E, Marin M, Diaz MS. *Pseudomonas aeruginosa*: a survey of resistance in 136 hospitals in Spain. The Spanish *Pseudomonas aeruginosa* Study Group. *Antimicrob Agents Chemother.* 1999;43(4):981-2.
28. Bonfiglio G, Carciotto V, Russo G, Stefani S, Schito GC, Debbia E, et al. Antibiotic resistance in *Pseudomonas aeruginosa*: an Italian survey. *J Antimicrob Chemother.* 1998;41(2):307-10.
29. Jones RN. Resistance patterns among nosocomial pathogens: trends over the past few years. *Chest.* 2001;119 (2 Suppl):397S-404S.
30. Haghi M, Maadi H, Delshad R, Nezhady MAM, Golizade SS. Antibiotic resistance pattern of *E coli*, *Staphylococcus aureus* and *Pseudomonas aeruginosa* isolated from burnt patients Urmia, Iran. *International Journal of Academic Research.* 2010;2(6 Pt 2): 377-80.
31. Zhang YP. Common pathogens in burn infections and changes in their drug sensitivity. *Zhonghua Zheng Xing Shao Shang Wai Ke Za Zhi.* 1991;7(2):108-10. [Article in Chinese]
32. Atoyebi OA, Sowemimo GO, Odugbemi T. Bacterial flora of burn wounds in lagos, Nigeria: a prospective study. *Burns.* 1992;18(6):448-51.
33. Vázquez F, Mendoza MC, Villar MH, Vindel A, Méndez FJ. Characteristics of *Pseudomonas aeruginosa* strains causing septicemia in a Spanish hospital 1981-1990. *Eur J Clin Microbiol Infect Dis.* 1992;11(8):698-703.
34. de Freitas, Barth AL. Antibiotic resistance and molecular typing of *Pseudomonas aeruginosa*: focus on imipenem. *Braz J Infect Dis.* 2002;6(1):1-7.
35. Nikbin VS, Abdi-Ali A, Feizabadi MM, Gharavi S. Pulsed field gel electrophoresis & plasmid profile of *Pseudomonas aeruginosa* at two hospitals in Tehran, Iran. *Indian J Med Res.* 2007;126(2):146-51.
36. Adel KK, Sabiha SS. Genetic site determination of antibiotic resistance genes in *Pseudomonas aeruginosa* by genetic transformation. *British Journal of Pharmacology and Toxicology.* 2010;1(2): 85-9.

پوخته

به‌لافوبون وهندك جوری دژه زینده هی به‌کتریای *Pseudomonas aeruginosa* ژنه‌خوشیت سوتنی ل نه‌خوشخانا سوتنا ل باریزگه‌ها دهوکی

پیشه‌کی و ئارمانج: ئه‌ف به‌کتریایه *Pseudomonas aeruginosa* دهیته هژمارتن کو دبیته هویئ توش بونا نه‌خوشیئت دری‌ژخایه یین وئیشین دژوار ژوان نه‌خوشییت به‌رگاریا له‌شی کیم لده‌ف وان، ونه‌خوشییت سوتنی، و ئه‌ف به‌کتریایه به‌رگاریی ل خودکه‌ت بشیویه کی سه‌ره کی ژ هنده‌ک دژه زینده‌هی antibiotic کوب زه‌حه‌ت بهیته چاره‌سه‌رکرن بو وان نه‌خوشیین توشی‌قی جور به‌کتریایی بن. ئارمانج ژ‌قی خاندنی ئه‌وه‌کو هه‌بونا‌قی جورئ به‌کتریایی ژ نه‌خوشیین سوتنی یا‌هه‌ی، ماوه‌یه‌کی هه‌ستیار وهیلا به‌رگاریی بو‌فان جورا وهنده‌ک دژه زینده‌هی کوده‌یته هژمارتن وه‌ک چاره‌سه‌ری بو‌فان نه‌خوشییت توژی‌قی جور به‌کتریایی دبن.

ریکین‌ف‌کولینی: د‌قی‌ف کولینی دا دیاریی 159 سامبل هاتنه‌کوم کرن یان دیارکرن دماوی ته‌موز -تشرینی یه‌که‌م 2010 ژنه‌خوشییت سوتنی ل نه‌خوشخانا سوتنا ل باریزگه‌ها دهوکی. و ئه‌ف سامبل هاتنه‌کوم کرن به‌ریکا بارچه‌به‌میبی یه‌کی باقژ ژهنده‌ک جهیت برینیت سوتنی دبه‌ن ژ نه‌خوشییت سوتنی دی جینین ل هنده‌ک شوینیت چاندنی ود که‌ینه‌حاجنی دا ماوی 24-48 ده‌مژمیرا ل تا‌قی‌گه‌ها نه‌خوشخانا سوتنی، و پاشتی‌قی چ‌ه‌ندی دی‌فان جوری سامبلا به‌ش به‌ش که‌ین بشیویه کی هه‌ره‌مه‌کی. پاشتی‌هنگی دی‌پاشکینا هه‌ستیار زینده‌کی، و ئینانا جوریت به‌رگاریی ب کارئینین disk diffusion.

نه‌جام: هژمارا سامبله 116، 159 سامبله به‌کتریا دیاریین، دنا‌ف برا وان 76 (47.7%) *Pseudomonas aeruginosa*، 20 *Klebsiella pneumoniae* (12.5%)، 10 *Escherichia coli* (6.2%)، 8 *Staphylococcus aureus* (5%)، 2 *Staphylococcus epidermidis* (1.2%) و 27 (43%) چ‌ه‌ نه‌جاما ناده‌ته‌مه‌ه. ئه‌جام هاتنه‌دیارکرن کو ری‌ژا *Pseudomonas aeruginosa* پ‌تربی ژ هه‌می جوریت دی‌ین بکتریا و ری‌ژه‌کا کیم *Escherichia coli*، *Staphylococcus aureus* و *Staphylococcus epidermidis* ژ نه‌خوشیین توشی‌فان به‌کتریا یا بیین. پاشکینا هه‌ستیار ژ جوری *Pseudomonas aeruginosa* دهیته وه‌رگرتن ژسوتین توشی‌قی به‌کتریایی دبن هه‌ستیاری پ‌تر کیمه‌ *amikacin*، *piperacillin*، *ciprofloxacin*، *imipenem*، *gentamicin*، *amoxi-clav*، *tetracycline*، *cef-taxime* و *vancomycin* د‌قی‌جور به‌کیریایی دا. وهاته‌دیارکرن ل پاشکینا جوری به‌رگاریی 12 به‌رگاریا جیاواز و جوری به‌ره‌لا‌ف به‌رگاریا 5 و 11.

ده‌ره‌نه‌جام: دیاریی د‌خاندنی دا کوری‌ژه‌یه‌کا زیده *Pseudomonas aeruginosa* یا‌هه‌ی ژنه‌خوشییت سوتنی یت کو توشی‌قی جور به‌کتریایی بیین و پ‌تریا‌فان جور به‌کتریا دیاردبن کو به‌رگاریی ژخودکه‌ن ژ دژه زینده‌هییا و دیاریی جورین به‌رگاریین جیاواز.

الخلاصة

نسبة تواجد وأنماط المقاومة المضادات الحيوية لبكتريا الزوائف الزنجارية المعزولة من أصابات الحروق في مستشفى الحروق في مدينة دهوك

خلفية واهداف البحث: تعتبر *Pseudomonas aeruginosa* بكتريا انتهازية تسبب إصابات مرضية حادة ومزمنة في مرضى ذات المناعة القليلة ومرضى القسطرة وكذلك مرضى الحروق. هذه البكتريا بصورة رئيسية مقاومة لعدد كبير من المضادات الحيوية، مما يصعب علاج الحالات المصابة بهذه البكتريا. الهدف من هذه الدراسة هو لايجاد نسبة تواجد بكتريا *Pseudomonas aeruginosa* في اصابات التهابات الحروق، وتحديد مدى حساسية هذه العزلات لمعظم المضادات الحيوية المتداولة، وكذلك اظهار انماط المقاومة لهذه العزلات.

طرق البحث: تضمنت هذه الدراسة 159 عينة جُمعت من الاصابات البكتيرية لالتهابات الحروق في مستشفى الحروق/مدينة دهوك، في الفترة الزمنية بين تموز - كانون الاول 2010. وتم جمع العينات باستخدام مسحات قطنية معقمة وأخذ عينات من القيح من إصابات الحروق وزرعها في الاوساط الزرعية التشخيصية والتحضين لمدة 24-48 ساعة في قسم المختبرات في مستشفى الحروق، وتم تشخيص العزلات باجراء الاختبارات التشخيصية الروتينية. تم إخضاع العزلات إلى إختبار فحص الحساسية للمضادات الحيوية باستخدام طريقة إنتشار القرص، وإيضاً إيجاد الانماط المقاومة للعزلات.

النتائج: العدد الكلي للعينات 159 عينة، 116 عينة اظهرت زرع بكتيري، من ضمنها 76 (47.7%) عَزلة *Pseudomonas aeruginosa* و 20 (12.5%) عَزلة *Klebsiella pneumoniae*، 10 (6.2%) *Escherichia coli*، 8 (5%) *Staphylococcus aureus*، 2 (1.2%) *Staphylococcus epidermidis* و 43 (27%) لم يعطى زرعاً بكتيرياً. أوضحت النتائج أن نسبة *Pseudomonas aeruginosa* كان الأكثر من بين المجاميع البكتيرية في اصابات الحروق، والنسبة الأقل للاصابات البكتيرية في الحروق كانت *Escherichia coli*، *Staphylococcus aureus* و *Staphylococcus epidermidis*. أُجري إختبار الحساسية لعزلات *Pseudomonas aeruginosa* المأخوذة من الاصابات البكتيرية لالتهابات الحروق حيث كانت أكثر حساسية للمضادات الحيوية imipenem تليها piperacillin، ciprofloxacin و amikacin بينما اظهرت أقل نسبة حساسية للمضادات الحيوية ceftaxime تليها tetracycline، gentamicin، cefotaxime، amoxiclav و vancomycin. كذلك تم إجراء تحليل انماط المقاومة للمضادات الحيوية حيث ظهرت 12 نمط مقاوم مختلف، وأن الانماط السائدة كانت نمط المقاومة 5 و 11.

الاستنتاجات: اظهرت هذه الدراسة زيادة في نسبة وجود *Pseudomonas aeruginosa* في الاصابات البكتيرية لالتهابات مرضى الحروق وأكثر هذه العزلات اظهرت مقاومة متعددة للمضادات الحيوية وكذلك اظهرت انواع مختلفة من نمط المقاومة.

BIOFILM FORMATION BY METHICILLIN RESISTANT STAPHYLOCOCCUS AUREUS (MRSA) WITHIN HOSPITAL AND COMMUNITY ACQUIRED URINARY TRACT INFECTIONS

AZA B. TAHA, PhD*

SABRIA M. SAID AL-SALIHI, PhD **

Submitted 10 Aug 2011; accepted 5 Jun 2012

ABSTRACT

Background and objectives Methicillin resistant *Staphylococcus aureus* is a significant cause of life-threatening human infections, which can switch from planktonic forms (i.e. single cells) to biofilms. Biofilm formation was often lower susceptibility to antibiotic treatments and development of chronic infections. The study was investigated biofilm formation by methicillin resistant *Staphylococcus aureus* isolated from patients with urinary tract infection. Meanwhile assess the relationship between biofilm formation and antibiotic resistance.

Methods *Staphylococcus aureus* were isolated and identification by standard methods from urinary tract infections at three teaching hospitals in Erbil city. Methicillin resistant *Staphylococcus aureus* were detected by PBP2a. Heterogeneity of methicillin resistant *Staphylococcus aureus* was determined by efficiency of plating method. Minimum inhibitory concentration of antibiotics was determined by agar dilution method. Biofilm forming ability of methicillin resistant *Staphylococcus aureus* was investigated.

Results Methicillin resistant *Staphylococcus aureus* were resistance to 10.92 ± 3.17 antibiotics. The percentage of biofilm formation by methicillin resistant *Staphylococcus aureus* was 82%. Strong biofilm formations were resistance to 13.40 ± 2.51 antibiotics, which is statistically higher than biofilm negative (Mean \pm SD = 6.56 ± 1.51).

Conclusions Most methicillin resistant *Staphylococcus aureus* were biofilm forming. Biofilm formation was correlated with multiple antibiotics resistance and to heterogeneous of methicillin resistant *Staphylococcus aureus*.

Duhok Med J 2012;6(1): 17-27.

Key words: Biofilm, MRSA, *Staphylococcus aureus*, Antibiotics, UTI, Infection

Biofilm is a multicellular creature made up of bacteria that irreversibly attached to a surface or interface, embedded in a matrix of extracellular polymeric substances.^{1,2} Biofilm formation is a complex developmental process involving attachment and immobilization on a surface, cell-to-cell interaction, microcolony formation, formation of a confluent biofilm, and development of a three dimensional biofilm structure.^{3,4}

Biofilms are responsible for several chronic diseases and show much greater resistance to antibiotics than their free-living counterparts.⁵ Bacterial cells within biofilms are inherently resistant to antimicrobial treatment and are difficult to eradicate from the infected individual. The high rates of morbidity and mortality associated with these infections.⁶ Biofilms are very hard to eradicate and responsible for a significant number of nosocomial and

* Lecturer, Head of Basic Sciences Department, College of Nursing, Hawler Medical University

** Professor of Medical Microbiology, Ministry of Higher Education and Scientific Research (KRG), Scientific Affairs

indwelling device-associated infections.⁷⁻⁹ Biofilms are notoriously difficult to eradicate and are a source of many chronic infections. According to the National Institutes of Health, biofilms are medically important, accounting for over 80% of microbial infections in the body.⁴

For decades, *Staphylococcus aureus* has been recognized as an important cause of disease around the world. It has become a major pathogen causing hospital and community acquired infections.^{10,11} *Staphylococcus aureus* and methicillin resistant *Staphylococcus aureus* (MRSA) are known to form biofilms on a variety of materials.¹² They can persist in clinical settings and gain increased resistance to antimicrobial agents through biofilm formation, which appears to be a bacterial survival strategy. Therefore, biofilms formed by MRSA have become resistant to the majority of antimicrobial agents. Due to gained multi-resistance, infections caused by MRSA are very difficult to treat.¹⁰ Factors contributing to the occurrence of MRSA infections are cross transmission via the hands of healthcare workers and high selective pressure exerted by broad-spectrum antibiotic therapy,¹¹ which is becoming increasingly difficult to treat given the ever-increasing incidence of MRSA and, more recently, the emergence of glycopeptide resistance.¹³

MRSA has emerged as a major clinical and epidemiological problem in hospitals. A distinctive feature of MRSA strains is their resistance not only to all β -lactam antibiotics, but also to a wide range of other antimicrobials, which makes MRSA infections difficult to manage and costly to treat.¹⁴ In recent years, the increasing incidence of urinary tract infections (UTI) caused by *Staphylococcus aureus* has been noted at the urology ward. The incidence of MRSA induced UTI is getting higher these days, especially for inpatients with an indwelling urinary catheter or those who are immunocompromised.^{10,15}

METHODS

Midstream urine samples were aseptically collected then cultured on blood agar (Oxoid, England) and mannitol salt agar (Oxoid, England) by standard techniques as soon as possible^{16,17} from 1367 patients with UTIs had documented pyuria (WBC>5/hpf) at Maternity, Rizgary and Hawler teaching hospitals in Erbil city from June 2010 through April 2011. Data information's were collected from patients, which included age, sex, and urinary catheterization. The infections were classified into community or hospital acquired infections based on Centers for Disease Control and Prevention definitions.¹⁸ MRSA infections in which MRSA was recognized in patients after 72 hours of hospitalization were considered as hospital acquired MRSA, this is the usual accepted duration of hospitalization required to develop the hospital-acquired infections. MRSA infections in outpatients were considered as community acquired MRSA. Exclusion criteria was pregnant woman, age less than 18 years, patient's hospitalizations less than 72 hours and outpatients have been hospitalizations in the last 21 days.

Staphylococcus aureus was identified by colony morphology, Gram staining, fermentation of mannitol, tube coagulase test, and Avipath Staph (Omega, UK).¹⁹ MRSA was detected using PBP2a kit (Oxoid, Japan) for detection PBP2a on cell wall of MRSA, which performed by sufficient colonies of *Staphylococcus aureus* suspended in 200 μ l extraction reagent 1 and heated in boiling water for 3 minutes. Tubes were cooled and 50 μ l extraction reagent 2 was added. Tubes were centrifuged at 1500xg for five minutes. Fifty microliters of suspension was mixed with 50 μ l sensitized latex suspension and rotated manually for 3 minutes while looking for agglutination, i.e., MRSA positive.^{20,21} Heterogeneous of MRSA was determined quantitatively by efficiency of plating method.²²

Overnight cultures of MRSA isolates were grown at 37°C in brain heart infusion broth (HiMedia, India) supplemented with 2% glucose and 2% sucrose. The culture was diluted 1:100 in brain heart infusion, and MRSA suspensions (200µL) were transferred to individual wells of a flat-bottom 96-well microplates (Costar, USA). After 48 hours at 37°C without shaking, wells were gently washed three times with distilled water, dried in an inverted position, and stained with 300µL of 2% crystal violet solution in water for 45 min. After staining, plates were washed 3 times with distilled water, and destained with 200µL of ethanol/acetone (95:5, vol/vol). A total of 200µL from each well was transferred to a new microplates, and analysis at optical density (OD) of 570 nm. Each assay was performed in triplicate, and the mean OD₅₇₀ value of tested wells was applied to biofilm forming ability. As a control, uninoculated medium was used to determine background OD. The mean OD₅₇₀ value from the control wells was subtracted from the mean OD₅₇₀ value of tested wells. The biofilm formation was divided into strong (OD₅₇₀ ≥0.5), medium (OD₅₇₀ ≥0.2 to <0.5), weak (OD₅₇₀ 0 to <0.2), and negative biofilm formation.^{23,24}

The minimum inhibitory concentration (MIC) was determined by the agar dilution method according to EUCAST (2001),²⁵ and BSAC (2010)²⁶ guidelines for Penicillin G (Sigma-Aldrich), Cefotaxime (Sigma-Aldrich), Ceftriaxone (Mepha), Cefepime (Exir), Tetracycline (Sigma-Aldrich), Doxycycline (Sigma-Aldrich), Amikacin (Sigma-Aldrich), Tobramycin (Sigma-Aldrich), Erythromycin (Sigma-Aldrich), Azithromycin (Fluka), Clarithromycin (Sigma-Aldrich), Ciprofloxacin (Fluka), Gatifloxacin (Cipla), Levofloxacin (Sigma-Aldrich), Moxifloxacin (Bayer), Ofloxacin (Sigma-Aldrich), Clindamycin (Sigma-Aldrich), Rifampicin (Sigma-Aldrich), and Chloramphenicol (Sigma-Aldrich). The bacteria were classified as

susceptible or resistant according to BSAC breakpoint criteria.²⁶

All statistical analyses were performed by Statistical Package for Social Sciences (SPSS). Descriptive statistics were given as arithmetic mean ± SD (standard deviation) and t-test. Comparisons between different groups were evaluated by one-way ANOVA with Duncan test at p<0.05. Correlation analyses were used to assess the relationship between two variables. All results were considered statically significant at the p<0.05 level.

RESULTS

Overall 157 (11.49%) *Staphylococcus aureus* isolates from 1367 UTIs were isolated; the prevalence of MRSA was 31.85% (50 of 157 *Staphylococcus aureus*) that isolated from 27 (54%) females and 23 (46%) males. The mean age of females (52.44±10.65) was statistically older than males (46.30±10.55) (Table 1). Statistical comparison was done by using Duncan test at P<0.05 for multiple comparison of 20 antibiotics revealed that the lower mean of MIC±SD were 0.09±0.06, 0.83±0.58, 0.96±0.14 and 0.96±0.57 µg/ml for Rifampin, Clarithromycin, Gatifloxacin and Clindamycin, respectively. However, significant differences were observed between MIC hospital and community acquired MRSA for Ceftriaxone, Amikacin, Azithromycin, Gatifloxacin and Clindamycin (Table 2).

Table 1. Gender and age of UTI patients with MRSA

Gender	No. (%)	Age (years)		
		Mean±SD	Min	Max
Female	27 (54)	52.44±10.65	24	64
Male	23 (46)	46.30±10.55	23	68
Total	50 (100)	49.62±10.94	23	68

Significant (t-value = 2.04, p-value = 0.047).

Min: Minimum; Max: Maximum

Table 2. MIC and antibiotics resistances of 34 hospital and 16 community acquired MRSA

Antibiotic	Antibiotic MIC					Antibiotic resistance		
	Mean±SD of MIC (mg/L)		Statistical analysis		Total Mean±SD (mg/L)	Hospital acquired	Community acquired	Total
	Hospital acquired	Community acquired	t-value	p-value		No. (%)	No. (%)	No.(%)
Penicillin G	70.59±25.86	58.00±31.39	1.50	0.14	66.56±28.06 ^h	34(100)	16(100)	50(100)
Cefotaxime	56.59±20.60	60.13±28.04	0.50	0.62	57.72±23.01 ^g	30(88.24)	15(93.75)	45(90)
Ceftriaxone	107.35±39.92	72.00±29.79	3.15	<0.01	96.04±40.28 ⁱ	32(94.12)	16(100)	48(96)
Cefepime	13.53±4.70	14.50±7.85	0.55	0.59	13.84±5.83 ^e	32(94.12)	16(100)	48(96)
Tetracycline	10.65±7.09	10.38±9.51	0.11	0.91	10.56±7.84 ^{de}	23(67.65)	9(56.25)	32(64)
Doxycycline	3.82±3.55	4.47±4.52	0.55	0.59	4.03±3.85 ^{abc}	14(41.18)	9(56.25)	23(46)
Gentamicin	10.71±6.74	9.41±7.74	0.61	0.55	10.29±7.02 ^{de}	28(82.35)	11(68.75)	39(78)
Amikacin	24.82±12.20	16.50±14.21	2.13	0.04	22.16±13.32 ^f	25(73.53)	7(43.75)	32(64)
Tobramycin	1.97±2.55	1.50±1.91	0.66	0.52	1.82±2.35 ^{ab}	5(14.71)	2(12.50)	7(14)
Erythromycin	6.43±4.17	9.53±21.29	0.58	0.57	7.42±12.35 ^{bcd}	29(85.29)	11(68.75)	40(80)
Azithromycin	12.38±6.17	6.69±8.81	2.65	0.01	10.56±7.52 ^e	29(85.29)	9(56.25)	38(76)
Clarithromycin	0.79±0.29	0.91±0.94	0.46	0.65	0.83±0.58 ^a	22(64.71)	7(43.75)	29(58)
Ciprofloxacin	5.85±5.66	5.38±10.54	0.21	0.84	5.70±7.46 ^{abcd}	25(73.53)	4(25.00)	29(58)
Gatifloxacin	0.94±0.16	1.00±0.00	2.10	0.04	0.96±0.14 ^a	0(0.00)	0(0.00)	0(0.00)
Levofloxacin	2.24±1.50	2.25±2.29	0.03	0.98	2.24±1.77 ^{ab}	2(5.88)	2(12.50)	4(8)
Moxifloxacin	1.21±0.76	1.56±2.52	0.55	0.59	1.32±1.54 ^a	4(11.76)	2(12.50)	6(12)
Ofloxacin	3.29±2.05	2.94±2.38	0.54	0.59	3.18±2.14 ^{ab}	16(47.06)	7(43.75)	23(46)
Clindamycin	1.09±0.60	0.69±0.40	2.79	0.01	0.96±0.57 ^a	22(64.71)	4(25.00)	26(52)
Rifampicin	0.10±0.06	0.08±0.06	0.85	0.40	0.09±0.06 ^a	14(41.18)	2(12.50)	16(32)
Chloramphenicol	10.35±4.42	8.25±4.95	1.51	0.14	9.68±4.65 ^{cde}	12(35.29)	4(25.00)	16(32)

The same letters mean no significant difference, the different letter mean significant difference at p<0.05.

Numbers of antibiotics resistance (Mean±SD) of MRSA were 10.92±3.17, which is higher in hospital acquired infection (11.59±2.69) than community acquired infection (9.50±3.71) (Table 3). The percentage of urinary catheterized of patients with MRSA infection were 66% that resistance to 11.79±2.46 antibiotics, which is statistically higher than non-catheterized patients (Mean±SD = 9.24±3.75) (Table 4). Among all MRSA isolated, 24% were heterogeneous MRSA and 76% were homogeneous MRSA. The numbers of antibiotics resistance (Mean±SD) of homogeneous MRSA (12.21±2.35) were statistically higher than heterogeneous MRSA (6.83±1.47) (Table 5).

Table 3. Multiple antibiotics resistance of MRSA isolated from patients with hospital and community acquired UTI

MRSA	No. (%)	Multiple antibiotics resistance (Mean±SD)
Hospital acquired	34 (68)	11.59±2.69
Community acquired	16 (32)	9.50±3.71
Total	50 (100)	10.92±3.17

Significant (*t*-value = 2.26, *p*-value = 0.02).

Table 4. Catheterization and numbers of antibiotics resistance of MRSA isolated from UTI

Urinary catheterization	No. (%)	Multiple antibiotics resistance (Mean±SD)
Catheterization	33 (66)	11.79±2.46
Non-catheterization	17 (34)	9.24±3.75
Total	50 (100)	10.92±3.17

Significant (*t*-value = 2.89, *p*-value = 0.006).

Table 5. Heterogeneous and homogeneous MRSA and multiple antibiotics resistance of MRSA isolates from UTI

MRSA	No. (%)	Multiple antibiotics resistance (Mean±SD)
Heterogeneous	12 (24)	6.83±1.47
Homogeneous	38 (76)	12.21±2.35
Total	50 (100)	10.92±3.17

Highly significant (*t*-value = 7.45, *p*-value <0.001).

Among 50 MRSA, 41 (82%) were biofilm formation of which 10%, 30%, 42% and 18% exhibited strong, medium, weak, and negative biofilm formation, respectively. Statistically multiple antibiotics resistance of strong biofilm formation MRSA were higher than medium, weak and negative biofilm formation (Table 6). Results found that correlations between: (i) biofilm formation by MRSA with multiple antibiotics resistance, (ii) biofilm formation and heterogeneous MRSA, (iii) biofilm formation and catheterization, (iv) multiple antibiotics resistance of MRSA and homogeneous MRSA, (v) multiple antibiotics resistance of MRSA and catheterization, and (vi) homogeneous MRSA and catheterization (Table 7).

Table 6. Biofilm formation by MRSA and numbers of antibiotics resistance

Biofilm formation	No. (%)	Multiple antibiotics resistance (Mean±SD)
Strong biofilm	5 (10)	13.40±2.51 ^a
Medium biofilm	15 (30)	11.93±1.49 ^b
Weak biofilm	21 (42)	11.48±3.11 ^b
Biofilm negative	9 (18)	6.56±1.51 ^b
Total	50 (100)	10.92±3.17

The same letters mean no significant difference, the different letter mean significant difference at *p*<0.05.

Table 7. Correlations of biofilm formation, multiple antibiotics resistance, homogeneous MRSA and catheterization of MRSA

Factors		Biofilm formation	Multiple antibiotics resistance	Homogeneous MRSA	Catheterization
Biofilm formation	Correlation	1.00	0.57*	0.58*	0.45*
	Significant	.	<0.001	<0.001	<0.001
Multiple antibiotics resistance	Correlation	0.57*	1.00	0.73*	0.39*
	Significant	<0.001	.	<0.001	0.01
Homogeneous MRSA	Correlation	0.58*	0.73*	1.00	0.39*
	Significant	<0.001	<0.001	.	0.01
Catheterization	Correlation	0.45*	0.39*	0.39*	1.00
	Significant	<0.001	0.01	0.01	.

* Correlation is significant at the 0.01 level (2-tailed).

DISCUSSION

Microbial biofilms have been associated with a variety of persistent infections, which respond poorly to conventional antibiotic therapy. This also helps in the spread of antibiotic resistant traits in nosocomial pathogens by increasing mutation rates and by the exchange of genes, which are responsible for antibiotic resistance.²⁷ MRSA were responsible for a significant number of biofilms related infections.¹³ The study found that UTI associated with MRSA was high, which is in agreement with other studies,^{10,13,23} they found biofilm formation was quite common among clinical MRSA isolates. The reasons for that are MRSA strains colonizes in a hospital and is easily transmitted via skin contact among patients and hospital staff, also increasing number of patients with catheterization.

The increasing prevalence of antibiotic resistant bacteria in hospitals and the community has significantly limited the effectiveness of current drugs resulting in treatment failure.^{28,29} In this study, multiple antibiotics resistance among MRSA has been reported that agreement with other studies.³⁰⁻³⁵ In addition, biofilm

forming by MRSA was correlation with increased antibiotics resistant among MRSA, which is in harmony with other study.^{36,37} Such biofilms are responsible for chronic UTIs, which are difficult to treat and show much greater resistance to antibiotics than their planktonic form counterparts.

Complicated urinary tract infection is often refractory to antimicrobial treatment. One of the reasons for this is the fact that the infection is a biofilm disease.³⁸ Biofilms have been shown to be up to 1000 times more resistant to antibiotics than planktonic cells of the same isolate.³⁹ The extracellular matrix of the biofilm is not only a passive diffusion barrier for antibiotics but is also actively shaped by species within the microbial biofilm communities, making biofilms extremely difficult to eradicate.^{31,40}

The results are similar to other studies,^{10,15} they found that significantly greater biofilm forming capacities of MRSA isolates from catheter related cases than of those from catheter unrelated cases. Indwelling medical device associated infections caused by *Staphylococcus aureus* biofilms are significant source of morbidity.⁸

REFERENCES

1. Supernak M, Świeczko-Żurek B. Reactions on the surface of the implant under the infection of biofilm. *Advances in Materials Science* 2010;10(4):1-7.
2. Donlan MR, Costerton JW. Biofilms: survival mechanisms of clinically relevant microorganisms. *Clin Microbiol Rev*. 2002;15(2):167-93.
3. Mohamed JA, Huang DB. Biofilm formation by enterococci. *J Med Microbiol*. 2007;56(Pt 12):1581-88.
4. Seidl K, Goerke C, Wolz C, Mack D, Berger-Bächi B, Bischoff M. *Staphylococcus aureus* CcpA affects biofilm formation. *Infect Immun*. 2008;76(5):2044-50.
5. Dugal S, Mamajiwala N. A novel strategy to control emerging drug resistant infections. *J Chem Pharm Res*. 2011;3(1):584-9.
6. Smith K, Perez A, Ramage G, Gemmelle CG, Lan C. Comparison of Biofilm-associated cell survival following in vitro exposure of methicillin-resistant *Staphylococcus aureus* biofilm to the antibiotics clindamycin, daptomycin, linezolid, tigecycline and vancomycin. *Int J Antimicrob Agents*. 2009;33(4):374-8.
7. Toté K, Berghe DV, Deschacht M, de Wit K, Maes L, Cos P. Inhibitory efficacy of various antibiotics on matrix and viable mass of *Staphylococcus aureus* and *Pseudomonas aeruginosa* biofilm. *Int J Antimicrob Agents*. 2009; 33(6):525-31.
8. Zhu Y, Weiss EC, Otto M, Fey PD, Smeltzer MS, Somerville GA. *Staphylococcus aureus* biofilm metabolism and the influence of arginine on polysaccharide intercellular adhesin synthesis, biofilm formation, and pathogenesis. *Infect Immun*. 2007;75(9):4219-26.
9. Donlan RM. Biofilms and device-associated infections. *Emerg Infect Dis*. 2001;7(5):277-81.
10. Ursic V, Tomic V, Kosnik M. Effect of different incubation atmospheres on the production of biofilm in methicillin-resistant *Staphylococcus aureus* (MRSA) grown in nutrient-limited medium. *Curr Microbiol*. 2008;57(4):386-90.
11. Alp E, Klaassen CH, Doganay M, Altoparlak U, Aydin K, Engin A, et al. MRSA genotypes in Turkey: persistence over 10 years of a single clone of ST239. *J Infect*. 2009;58(6):433-8.
12. O'Neill E, Pozzi C, Houston P, Humphreys H, Robinson DA, Loughman A, et al. A novel *Staphylococcus aureus* biofilm phenotype mediated by the fibronectin-binding proteins, FnBPA and FnBPB. *J Bacteriol*. 2008;190(11):3835-50.
13. O'Neill E, Pozzi C, Houston P, Smyth D, Humphreys H, Robinson DA, et al. Association between methicillin susceptibility and biofilm regulation in *Staphylococcus aureus* isolates from device-related infections. *J Clin Microbiol*. 2007;45(5):1379-88.
14. Petrelli D, Repetto A, D'Ercole S, Rombini S, Ripa S, Prenna M, et al. Analysis of methicillin-susceptible and methicillin-resistant biofilm-forming *Staphylococcus aureus* from catheter infections isolated in a large Italian hospital. *J Med Microbiol*. 2008; 57(Pt 3):364-72.
15. Araki M, Kariyama R, Monden K, Tsugawa M, Kumon H. Molecular epidemiological studies of *Staphylococcus aureus* in urinary tract infection. *J Infect Chemother*. 2002;8(2):168-74.
16. Alonto AM. Urinary tract infections. In: Mahon CR, Lehman DC, Manuselis G, editors. *Textbook of diagnostic microbiology*. 3rd ed. Philadelphia: Elsevier; 2007. p. 1020-2.
17. Engbaek K, El-Nageh MM, Groen J.

- Specimen collection and transport for microbiological investigation. Alexandria, Egypt: WHO; 1995.
18. Garner JS, Jarvis WR, Emori TG, Horan TC, Hughes JM. CDC definitions for nosocomial infections. *Am J Infect Control*. 1988;16:128-40.
 19. Harrison LS. Staphylococci. In: Mahon CR, Lehman DC, Manuselis G, editors. *Textbook of diagnostic microbiology*. 3rd ed. Philadelphia: Elsevier; 2007. p. 367-407.
 20. Brown DF, Edwards DI, Hawkey PM, Morrison D, Ridgway GL, Towner KJ, et al. Guidelines for the laboratory diagnosis and susceptibility testing of methicillin-resistant *Staphylococcus aureus* (MRSA). *J Antimicrob Chemother*. 2005;56(6):1000-18.
 21. Mohanasoundaram KM, Lalitha MK. Comparison of phenotypic versus genotypic methods in the detection of methicillin resistance in *Staphylococcus aureus*. *Indian J Med Res*. 2008;127:78-84.
 22. Fung-Tomc J, Huczko E, Gradelski E, Denbleyker K, Bonner DP, Kessler RE. Emergence of homogeneously methicillin-resistant *Staphylococcus aureus*. *J Clin Microbiol*. 1991; 29(12):2880-3.
 23. Ando E, Monden K, Mitsuhashi R, Kariyama R, Kumon H. Biofilm formation among methicillin-resistant *Staphylococcus aureus* isolates from patients with urinary tract infection. *Acta Medica Okayama*. 2004;58(4):207-14.
 24. Christensen GD, Simpson WA, Younger JJ, Baddour LM, Barrett FF, Melton DM, et al. Adherence of coagulase-negative staphylococci to plastic tissue culture plates: a quantitative model for the adherence of staphylococci to medical devices. *J Clin Microbiol*. 1985;22(6):996-1006.
 25. European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID). EUCAST Definitive Document E.DEF 3.1, June 2000: determination of minimum inhibitory concentrations (MICs) of antibacterial agents by agar dilution. *Clin Microbiol Infect*. 2001;6(9):509-15.
 26. British Society for Antimicrobial Chemotherapy. BSAC methods for antimicrobial susceptibility testing. Version 9.1 [Internet]. 2010 [cited 2010 Dec 12]. Available from: <http://www.bsac.org.uk>
 27. Bose S, Ghosh AK. Biofilms. *Journal of Clinical and Diagnostic Research*. 2011;5(1):127-30.
 28. Nwanze PI, Nwaru LM, Oranusi S, Dimkpa U, Okwu MU, Babatunde BB, et al. Urinary tract infection in Okada village: Prevalence and antimicrobial susceptibility pattern. *Sci Res Essays*. 2007;2(4):112-6.
 29. Dugal S, Mamajiwala N. A novel strategy to control emerging drug resistant infections. *J Chem Pharm Res*. 2011;3(1):584-9.
 30. Pillar MC, Draghi DC, Sheehan DJ, Sahm DF. Prevalence of multidrug-resistant, methicillin-resistant *Staphylococcus aureus* in the United States: findings of the stratified analysis of the 2004 to 2005 LEADER Surveillance Programs. *Diagn Microbiol Infect Dis*. 2008;60: 221-4.
 31. Borg MA, Kraker M, Scicluna E, Sande-Bruinsma N, Tiemersma E, Monen J et al. Prevalence of methicillin-resistant *Staphylococcus aureus* (MRSA) in invasive isolates from southern and eastern Mediterranean countries. *J Antimicrob Chemother*. 2007;60:1310-5.
 32. Martins A, Couto I, Aagaard L, Martins M, Viveiros M, Kristiansen JE et al. Prolonged exposure of methicillin-resistant *Staphylococcus aureus* (MRSA) COL strain to increasing concentrations of oxacillin results in a multidrug-resistant phenotype. *Int J Antimicrob Agents*. 2007;29:302-5.

33. Rajaduraipandi K, Mani KR, Panneerselvam K, Mani M, Bhaskar M, Manikandan P. Prevalence and Antimicrobial Susceptibility Pattern of Methicillin Resistant *Staphylococcus aureus*: A Multicentre Study. Indian J Med Microbiol. 2006;24(1):34-8.
34. Shittu AO, Lin J. Antimicrobial susceptibility patterns and characterization of clinical isolates of *Staphylococcus aureus* in KwaZulu Natal province, South Africa. BMC Infect Dis. 2006;6:125- 37.
35. Onanuga A, Oyi AR, Onaolapo JA. Prevalence and susceptibility pattern of methicillin resistant *Staphylococcus aureus* isolates among healthy women in Zaria, Nigeria. Afr J Biotechnol. 2005;4(11):1321-4.
36. Lewis K. Riddle of biofilm resistance. Antimicrob Agents Chemother. 2001;45(4): 999-1007.
37. Estrela AB, Abraham W. Combining biofilm-controlling compounds and antibiotics as a promising new way to control biofilm infections. Pharmaceuticals. 2010;3(5):1374-93.
38. Tsukamoto T, Matsukawa M, Sano M, Takahashi S, Hotta H, Itoh N, et al. Biofilm in complicated urinary tract infection. Int J Antimicrob Agents. 1999;11(3-4):233-6.
39. Fricks-Lima J, Hendrickson CM, Allgaier M, Zhuo H, Wiener-Kronish JP, Lynch SV, et al. Differences in biofilm formation and antimicrobial resistance of *Pseudomonas aeruginosa* isolated from airways of mechanically ventilated patients and cystic fibrosis patients. Int J Antimicrob Agents. 2011;37(4):309-15.
40. Shafahi M, Vafai K. Synthesis of biofilm resistance characteristics against antibiotics. Int J Heat Mass Transfer. 2010;53:2943-50.

پوختە

دروست بوونی biofilm بە ھۆی خڕۆکە ھیشوویە زیرینەکان بەرگریکەر لە مەسیسلین (MRSA) ی دەستکەوتوو
لە بۆری میزی تووشبووکان لە نەخۆشخانە و کۆمەلگە

پێشەکی و ئارمانج: خڕۆکە ھیشوویە زیرینەکان بەرگریکەر لە مەسیسلین (MRSA) بە ھۆیەکی گرتە داوە نریت بۆ تووش بوونی مرۆڤ و تیکدانێ زۆر. وە دەتوانی شێوھەکی بگۆری لە شێوھەکی تاک خانەیی (planktonic) بۆ شێوھەکی biofilm کە لە زۆریەکی کاتدا زۆر بە کەمی لە ناو دەبرێن بە دژە زیندەکان بە تاییەتی لە کاتی ھەوکردنی درژ خایەندا. ئەم توێژینە وە لە دروستبوونی biofilm ەکان لە خڕۆکە زیرینە بەرگریکەر لە مەسیسلین دەکۆڵێتە وە کە وەرگیراون لە نەخۆشخانە کە تووش بوون بە ھەوکردنی بۆری میز. لە ھەمان کاتیشدا بۆ ھەلسەنگاندن و دیاریکردنی پەیوەندنی لە نێوان biofilm ە دروستبووکان و دژە زیندە بەرگریکەرەکانیاندا.

ریکێن فەکولینی: لە سێ نەخۆشخانە فیکاری شارێ ھەولێر کارەکە ئەنجامدرا بە وەرگرتن و جیاکردنە وە ی خڕۆکە ھیشوویە زیرینەکان لە بۆری میزی تووشبووکان بە ھەوکردن ئێویش بە رێگە ی پێوانەکردن وە بە دیاریکردنی PBP2a. کە heterogeneous MRSA خڕۆکە ھیشوویە زیرینە بەرگریکەر لە مەسیسلین جیاواز دەرکەوتن، بە چالاکی رێگە ی رووکەش کردن (efficiency of plating). وە نزمترین ئاستی خەستی بۆ نەھیشتەنی دژە زیندەکان دیاری کرا بە رێگە ی ئاگاری روون، وە ھەروەھا biofilm ە دروستبووکانیش لە خڕۆکە ھیشوویە زیرینە بەرگریکەر لە مەسیسلین دیاری کرا.

ئەنجام: خڕۆکە ھیشوویە زیرینە بەرگریکەر لە مەسیسلین بەرگریکەر بوون بۆ $3,17 \pm 10,92$ لە دژە زیندەکان وە رێژە ی سەدی دروستبوونی biofilm ەکان لە خڕۆکە ھیشوویە زیرینە بەرگریکەر لە مەسیسلین 82% بوو. وە دروستبوونی biofilm ی بە ھێز کە بەرگریکەر بوون بۆ $2,51 \pm 13,40$ لە دژە زیندەکان کە ئەمەش رێژە یەکی بەرز بوو لە چاو دروست نەبوونی biofilm ەکان کە $(1,51 \pm 6,56 = SD \pm Mean)$.

دەرئەنجام: زۆریەکی خڕۆکە ھیشوویە زیرینە بەرگریکەر لە مەسیسلین لە شێوھەکی biofilm بوون، کە ئەم biofilm نەش پەیوەندبوون بە دژە زیندە جیاوازە بەرگریکەرەکانە وە، وە بە خڕۆکە ھیشوویە زیرینە بەرگریکەر لە مەسیسلین جیاوازەکانە وە.

الخلاصة

تشكيل biofilm في المكورات العنقودية الذهبية المقاومة للمثسلين (MRSA) المسببة لالتهاب المسالك البولية المكتسبة من عدوى المستشفى والمجتمع

خلفية وأهداف البحث: المكورات العنقودية الذهبية المقاومة للمثسلين (MRSA) هي سبب مهم في العدوى التي تهدد الحياة و التي يمكن ان تتحول من خلايا منفردة (planktonic) إلى biofilms. تشكيل biofilm غالبا ما تكون أقل استجابة للعلاج بالمضادات الحيوية وتسبب الالتهابات المزمنة. الدراسة تحققت من تشكيل biofilm في المكورات العنقودية الذهبية المقاومة للمثسلين المعزولة من المرضى الذين يعانون من التهاب المسالك البولية. في الوقت نفسه تم تقييم العلاقة بين تشكيل biofilm والمقاومة للمضادات الحيوية.

طرق البحث: تم عزل المكورات العنقودية الذهبية بواسطة الطرق القياسية من التهابات المسالك البولية في ثلاث مستشفيات تعليمية في مدينة أربيل. تم الكشف عن المكورات العنقودية الذهبية المقاومة للمثسلين بواسطة الكشف عن PBP2a. تم تحديد heterogeneous MRSA efficiency of plating. تم تحديد الحد الأدنى للتركيز المثبطة للمضادات الحيوية بطريقة تخفيف الأكار. تمت الكشف عن قدرة تشكيل biofilm في المكورات العنقودية الذهبية المقاومة للمثسلين.

النتائج: المكورات العنقودية الذهبية المقاومة للمثسلين مقاومة لـ $10,92 \pm 3,17$ من المضادات الحيوية. النسبة المئوية لتشكيل biofilm في المكورات العنقودية الذهبية المقاومة للمثسلين كانت 82%. تشكيل biofilm قوي مقاومة لـ $13,40 \pm 2,51$ من المضادات الحيوية وهي أعلى من الناحية الإحصائية في حال عدم تشكيلها لـ $6,56 \pm SD$ biofilm ($\pm 1,51$).

الاستنتاجات: معظم المكورات العنقودية الذهبية المقاومة للمثسلين كونت biofilm و تكوينها لـ biofilm ترتبط مع مقاومتها للمضادات الحيوية.

THE EFFECT OF *HELICOBACTER PYLORI* ERADICATION THERAPY ON
PLATELET COUNT IN IDIOPATHIC THROMBOCYTOPENIA: A PILOT STUDY

ASWAD AL-OBEIDY, FICMS(GE&H)*
AHMED Y. ELMESHEDANY, FIBMS (Clin Hematol)**

Submitted 16 Nov 2011; accepted 5 Jun 2012

ABSTRACT

Background and objectives Conflicting reports on the relationship between *Helicobacter pylori* infection and idiopathic thrombocytopenia had previously appeared in the literatures. This study examines the effect of *Helicobacter pylori* eradication on platelet counts in Iraqi patients with Idiopathic Thrombocytopenic Purpura (ITP).

Methods The study population comprised 31 Iraqi patients with chronic ITP and a platelet count of less than $100.0 \times 10^9/L$ and positive serum *H. pylori* antibodies (indirect immunofluorescence). They were divided into two groups, the first (17 patients) received anti *H. Pylori* plus conventional treatment for ITP, the second group (14 patients) received conventional treatment for ITP only. The effect of *H. pylori* eradication on platelet count was evaluated 6 months after therapy.

Results There was significant improvement in platelet count in response to conventional treatment in both groups but there were no significant improvement after *H. Pylori* eradication therapy.

Conclusions Based on this pilot study eradication of *H. pylori* does not appear to be effective in increasing platelet count in *H. pylori*-positive patients with chronic ITP.

Duhok Med J 2012;6(1): 28-33.

Key words: Eradication therapy, *H. pylori*, Immune thrombocytopenia

Helicobacter pylori is a slow-growing, microaerophilic, highly motile, gram-negative spiral bacterial organism etiologically linked to histologic gastritis, peptic ulcer disease, primary B cell gastric lymphoma, and adenocarcinoma of the stomach. A number of other conditions have been suggested as causally related to *H. pylori*, but the data supporting these associations are weak¹. Two conditions that have increasingly been associated with *H. pylori* and have been assessed by treatment trials are iron deficiency and idiopathic thrombocytopenic purpura²⁻⁴. Immune thrombocytopenic purpura is an acquired disorder leading to immune-mediated destruction of platelets and possibly inhibition of platelet release from the megakaryocyte. The presence of auto-

antibodies, often directed against platelet membrane glycoprotein IIb-IIIa, causes the premature removal of platelets by the monocyte-macrophage system. Occasionally, antigen-antibody immune complexes adhere to platelets at their F_c receptor, resulting in their premature removal from the circulation.⁵

The study which is a pilot study was aimed at evaluating the value of *H. pylori* eradication on the outcome of Management of ITP.

METHODS

This is a prospective study conducted in a period between December 2010 and October 2011 in Nanakelly hospital for blood diseases in Hawler, Iraq. During this

* Consultant Gastroenterologist

** Lecturer, Hawler Medical University-Medical college

Correspondence author: Ahmed Y. El-Meshedany. Email: dahmedk@yahoo.com, Mobile: 07702710298

period a total of 106 patients were diagnosed as Idiopathic thrombocytopenic Purpura (ITP). Out of these patients thirty one were *H. pylori* antibody positive. Only the latter group of patients were recruited for the purposes of this study. Eleven were males and twenty were females. Their mean age was 27.3 years (range 7-61). ITP was defined by idiopathic thrombocytopenia (platelets less than $100 \times 10^9/L$) when other causes had been excluded and with a normal active bone marrow. *H. pylori* infection was assessed by the detection of serum antibodies (indirect immunofluorescence).

The recruited patients were divided into two groups. The 1st group comprised 17 patients and received anti-*H. Pylori* eradication plus conventional treatment for ITP, the 2nd group (14 patients) received conventional treatment for ITP only. *H. pylori* eradication therapy consisted of amoxicillin (1000 mg twice daily), metronidazole (500 mg 3 times daily), and esomeprazole (40 mg twice daily) for 7 days. Conventional treatment for ITP included prednisolone 1 mg per Kg in cases where platelet count was below $30 \times 10^9/L$. Platelets counts were monitored every 2 weeks and counts at 6 months post therapy were taken.

Statistical analysis included using t – test. $P < 0.05$ was considered significant.

RESULTS

Out of the total of 106 Idiopathic thrombocytopenic patients, 31 (29.2%) were *H. pylori* antibody positive. The latter were divided into two groups, the 1st group (17 patients) who received anti *H. Pylori* plus conventional treatment had a mean \pm SD platelet count at diagnosis of $28.5 \pm 29.4 \times 10^9/L$ compared to $18.5 \pm 18.7 \times 10^9/L$ for 2nd group who only received conventional therapy ($p=0.28$).

Following therapy the platelets mean count rose to $107.1 \pm 57.2 \times 10^9/L$ for the 1st group and $128.6 \pm 82.9 \times 10^9/L$ for the

second group, which again was insignificant ($p=0.400$).

The mean increase in platelet count in group 1 was slightly lower at $80.2 \pm 68.4 \times 10^9/L$ compared to group 2, with a mean of $110.2 \pm 84.5 \times 10^9/L$, however this was not statistically significant ($p=0.27$).

DISCUSSION

Several lines of direct and indirect evidences suggest that infectious agents may influence the occurrence or the course of some auto-immune diseases.⁶⁻⁷

The mechanism by which *H. pylori* may play a role in ITP pathogenesis remains unclear. A chronic immunological stimulus induced by *H. pylori* or an immune mimicry between platelets and *H. pylori* antigens has been suggested as the cause of *H. pylori*-induced ITP.⁸ Although it has been demonstrated that antibodies against *H. pylori* cross-react with human tissues, such as gastric epithelial cells, ductal cells of salivary gland, and renal tubular cells,⁹ there is no conclusive support of cross-reactivity with platelets, and conflicting reports on its association with ITP.¹⁰⁻¹³

The prevalence of *H. pylori* among our ITP patients is comparable to previous studies, including that of Michel and coworkers, who reported a rate 21.6% among 74 patients 10 year or older in age with chronic ITP and platelets count $< 60 \times 10^9/L$.¹⁴

In the present study, we assessed the change in platelet count over time in two groups, there was significant improvement in platelet count in response to conventional treatment in both groups but there was no actual improvement after *H. Pylori* eradication. This result is consistent with that of Michel and colleagues who eradicated *H. pylori* in 14 of 15 patients but only one had significant transient platelets improvement.¹⁴ However, Our results is in contrast to those of Gasbarrini et al who documented *H. pylori* infection in

11 of 18 patients; 8 of the 11 patients in whom *H. pylori* was eradicated experienced significant platelet increments.¹¹ Similarly Emilia et al observed *H. pylori* in 13 of 30 patients with chronic ITP, and increased platelet counts occurred in 6 of 12 patients following eradication.¹⁵

H. pylori infection in particular has been recently under intensive clinical investigation. Interestingly, in many countries with a high prevalence of the infection, bacterial eradication reverses the thrombocytopenia in about 50% of cases with chronic ITP. The situation is different in North America, where *H. pylori* infection is found in a low proportion of cases and eradication seldom has any effect on the platelet count. A plausible explanation is that the *H. pylori* strains differ in different parts of the world.¹⁶ Other possibilities include variations in bacterial virulence factors, such as CagA expression or host immunological class II HLA factors.^{17,18} Most Japanese *H. pylori* strains are CagA-positive, unlike most American strains. Patients infected with CagA-positive strains (as measured by anti-CagA IgG antibodies) are thought to have increased platelet response to *H. pylori* eradication compared with those who are CagA-negative or who have low serum titres of CagA antibodies. Serum levels of anti-CagA IgG may be used as a predictor of response to *H. pylori* treatment.^{17,18}

In conclusion, the results in this preliminary study suggest similar to some previous reports, no significant additional benefits of *H. pylori* eradication in ITP. However, further studies including larger number of cases, as well as studies on the prevalence of *H. pylori* among patients with ITP is suggested.

REFERENCES

1. Leontiadis GI, Sharma VK, Howden CW. Non-gastrointestinal tract associations of *Helicobacter pylori*

- infection. Arch Intern Med. 1999;159(9):925-40.
2. Barabino A. *Helicobacter pylori*-related iron deficiency anemia: a review. Helicobacter. 2002;7(2):71-5.
3. Ando K, Shimamoto T, Tauchi T, Ito Y, Kuriyama Y, Gotoh A, et al. Can eradication therapy for *Helicobacter pylori* really improve the thrombocytopenia in idiopathic thrombocytopenic purpura? Our experience and a literature review. Int J Hematol. 2003;77(3):239-44.
4. Veneri D, Franchini M, Gottardi M, D'Adda M, Ambrosetti A, Krampera M, et al. Efficacy of *Helicobacter pylori* eradication in raising platelet count in adult patients with idiopathic thrombocytopenic purpura. Haematologica. 2002;87(11):1177-9.
5. Cines DB, McMillan R. Management of adult idiopathic thrombocytopenic purpura. Annu Rev Med. 2005;56:425-42.
6. Ernst PB, Jin Y, Reyes VE, Crowe SE. The role of the local immune response in the pathogenesis of peptic ulcer formation. Scand J Gastroenterol Suppl. 1994;205:22-8.
7. Bussel JB. Novel approaches to management of immune thrombocytopenic purpura: results of recent trials. Orlando: American Society of Hematology; 2001.
8. Gasbarrini A, Franceschi F. Autoimmune diseases and *Helicobacter pylori* infection. Biomed Pharmacother. 1999;53(5-6):223-6.
9. Ko GH, Park HB, Shin MK, Park CK, Lee JH, Youn HS, et al. Monoclonal antibodies against *Helicobacter pylori* cross-react with human tissue. Helicobacter. 1997;2(4):210-5.
10. Gasbarrini A, Franceschi F, Tartaglione R, Landolfi R, Pola P, Gasbarrini G. Regression of autoimmune thrombocytopenic purpura after eradication of *Helicobacter pylori* [letter]. Lancet. 1998;352:87-8.

11. Emilia G, Longo G, Luppi M, Gandini G, Morselli M, Ferrara L, et al. Helicobacter pylori eradication can induce platelet recovery in idiopathic thrombocytopenic purpura. *Blood*. 2001 ;97(3):812-4.
12. Veneri D, Franchini M, Gottardi M, D'Adda M, Ambrosetti A, Krampera M, et al. Efficacy of Helicobacter pylori eradication in raising platelet count in adult patients with idiopathic thrombocytopenic purpura. *Haematologica*. 2002;87(11):1177-9.
13. Kohda K, Kuga T, Kogawa K, Kanisawa Y, Koike K, Kuroiwa G, et al. Effect of Helicobacter pylori eradication on platelet recovery in Japanese patients with chronic idiopathic thrombocytopenic purpura and secondary autoimmune thrombocytopenic purpura. *Br J Haematol*. 2002;118(2):584-8.
14. Michel M, Khellaf M, Desforges L, Lee K, Schaeffer A, Godeau B, et al. Autoimmune thrombocytopenic purpura and Helicobacter pylori infection. *Arch Intern Med*. 2002;162(9):1033-6.
15. Stasi R, Sarpatwari A, Segal JB, Osborn J, Evangelista ML, Cooper N, et al. Effects of eradication of Helicobacter pylori infection in patients with immune thrombocytopenic purpura: a systematic review. *Blood*. 2009;113(6):1231-40. Epub 2008 Oct 22.
16. Gasbarrini A, Franceschi F, Does H. Pylori infection play a role in idiopathic thrombocytopenic purpura and in other autoimmune diseases? *Am J Gastroenterol*. 2005;100(6):1271-3.
17. Takahashi T, Yujiri T, Tanizawa Y. Helicobacter pylori and chronic ITP: the discrepancy in the clinical responses to eradication therapy might be due to differences in the bacterial strains. *Blood*. 2004;104(2):594.
18. Kodama M, Kitadai Y, Ito M, Kai H, Masuda H, Tanaka S, et al. Immune response to CagA protein is associated with improved platelet count after Helicobacter pylori eradication in patients with idiopathic thrombocytopenic purpura. *Helicobacter*. 2007;12(1):36-42.

پوخته

به کتريايي ئىچ پايلىرى و كه مېوونى خه پله خوینى ناديار و کاريگه رى له ناوبردنى به کتريايي ئىچ پايلىرى له سه ر ژماره يى
خه پله خوینه کان

پيشهكى و ئارمانج: په يوه ندى نيوان په تابوون به به کتريايي ئىچ پايلىرى و كه مېوونى خه پله خوینى ناديار باسكراوه له ليكولينه وه كانى پيشوو. له م ليكولينه وه يه دا جه خت له سه ر کاريگه رى له ناوبردنى به کتريايي ئىچ پايلىرى له سه ر زيادبوونى ژماره يى خه پله خوینه کان.

ريکتن فه کولينى: سى و يه ك نه خوشى توشبوو به نه خوشى كه مېوونى خه پله خوینى ناديارى دريژخايه ن هاتنه ديارى كرن كه هه موويان نمونه ي خوینيان پوزه تيفبوون بۆ به کتريايي ئىچ پايلىرى به ريگايى ره نگدانه وه ي ناراسته وخو (Indirect immunofluorescence). نه خوشه كان کران به دوو گرووپ، گرووپى يه كه م (17 نه خویش) هاتنه چاره سه ركردن به دژه به کتريايي ئىچ پايلىرى له گه ل چاره سه ركردن ته قلديى كه مېوونى خه پله خوینى دريژخايه ن، وه گرووپى دووهم (14 نه خویش) هاتنه چاره سه ركردن ته نها به ريگه چاره ي ته قلديى. له شه ش مانگ دا به دوا داچوونى حاله ته كان كرا.

ئه نجام: ئاماره كان ئه وه نيشان ده دن كه له ناوبردنى به کتريايي ئىچ پايلىرى رۆلېكى گرنكى نيه بۆ زيادكردن ژماره يى خه پله خوینه كان.

ده رئه نجام: له ناوبردنى به کتريايي ئىچ پايلىرى هېچ رۆلېكى نيه له به رزكردنه وه يى ئاستى خه پله خوینه كان له ئه و نه خویشانه يى كه توشبوون به كه مېوونى خه پله خوینى دريژخايه ن.

الخلاصة

تأثير استئصال جرثومة الهليكوباكتر على عدد الأقرص الدموية لدى المرضى المصابين بنقص الأقرص الدموية الفرغرية العفوي

خلفية وأهداف البحث: العلاقة بين الهليكوباكتر ونقص الأقرص الدموية الفرغرية العفوي مثبتة سابقاً. هذه الدراسة تحدد تأثير استئصال جرثومة الهليكوباكتر على عدد الأقرص الدموية.

طرق البحث: هذه الدراسة تتضمن واحد وثلاثون مريض عراقي مصابون بمرض نقص الأقرص الدموية الفرغرية العفوي مع عدد الأقرص أقل من 100000 مصابون بجرثومة الهليكوباكتر. المرضى قسموا إلى مجموعتين: المجموعة الأولى (17) استلمت علاج استئصال جرثومة الهليكوباكتر مع العلاج التقليدي لمرض نقص الأقرص الدموية الفرغرية. المجموعة الثانية استلمت العلاج التقليدي لمرض نقص الأقرص الدموية الفرغرية فقط. تأثير استئصال جرثومة الهليكوباكتر على عدد الأقرص الدموية تم تقييمه لمدة 6 أشهر.

النتائج: كان هناك تحسن معتبر بعدد الأقرص الدموية في المجموعتين رداً على العلاج التقليدي ولكن لم يكن هناك تحسن معتبر بعد استئصال جرثومة الهليكوباكتر.

الاستنتاجات: استئصال جرثومة الهليكوباكتر غير فعال في زيادة عدد الأقرص الدموية عند مرضى نقص الأقرص الدموية الفرغرية المصابون بجرثومة الهليكوباكتر.

**METHYLENETETRAHYDROFOLATE REDUCTASE (C677T) MUTATION IN
HEALTHY INDIVIDUALS IN ERBIL - IRAQ**

FARIDA FA. NERWEYI, PhD*

Submitted 22 Jan 2011; accepted 5 Jun 2012

ABSTRACT

Background and objectives Methylenetetrahydrofolate reductase is one of the main regulatory enzymes of homocysteine metabolism. A 677C→T mutation in the methylenetetrahydrofolate reductase gene results in decreased enzymatic activity, and contributes to increased plasma homocysteine. The association between the C677T mutation in the methylenetetrahydrofolate reductase gene and vascular disease is controversial, and may be affected by ethnic origin. The aim of this study was to study the frequency of the C677T methylenetetrahydrofolate reductase mutation in healthy individuals from Erbil-Iraq.

Methods A total of 100 healthy individuals attending the premarital screening center in Erbil city were recruited. methylenetetrahydrofolate reductase (C677T) gene polymorphism was investigated in all of them by the polymerase chain reaction and restriction fragment length polymorphism.

Results Methylenetetrahydrofolate reductase C677T was documented in homozygous and heterozygous state in 6% and 37% respectively.

Conclusions Methylenetetrahydrofolate reductase C677T mutation is commonly encountered among healthy individuals in Erbil city, although it was rather less frequent than that documented earlier in Duhok, to the north of the former city. The clinical implications of our finding require further clinical studies.

Duhok Med J 2012;6(1): 34-42.

Key words: C677T *MTHFR* gene mutation, PCR-RFLP, Duhok - Iraq

The methylenetetrahydrofolate reductase (MTHFR) gene, located on the short arm of chromosome 1 (1p36.3), presents two common polymorphisms involving nucleotides C677T and A1298C. The change of C for T at position 677 causes the substitution of alanine for valine in the MTHFR protein and the consequent reduction in enzyme activity. The specific activity of the MTHFR enzyme is reduced by 35% in the presence of heterozygous, genotype C/T, compared to the normal genotype C/C, and by 70% in homozygous, genotype T/T.¹ MTHFR is an enzyme in the transmethylation pathway where homocysteine (Hcy) is converted to methionine, thus impaired enzyme activity leads to hyperhomocysteinemia.² MTHFR enzyme has important role in metabolic pathway of folate and nucleotide methylation.³

Presence of T allele at position 677 of MTHFR gene leads to reduction of MTHFR activity and DNA hypomethylation. Genetic variation in this gene influences susceptibility to occlusive vascular disease, neural tube defects, colon cancer and acute leukemia, and mutations in this gene are associated with methylenetetrahydrofolate reductase deficiency.⁴

There is ethnic variability in the frequency of the T allele–frequency. The latter in Mediterranean/Hispanics is greater than the frequency in Caucasians which, in turn, is greater than in Africans/African-Americans.⁵ Mutations of the methylenetetrahydrofolate reductase (MTHFR) gene have been shown to be associated with a predisposition to developing diabetic nephropathy (DN) in specific populations.⁶ Elevated levels of

* Lecturer, Faculty of Science - Department of Scientific Research Center, University of Duhok. Email: fn964@yahoo.com; Tel: +9647504494508.

total plasma homocysteine have been linked to increased all-cause mortality,⁷ arteriosclerosis,⁸ and thromboembolism.^{9,10} This study was aimed to determine the prevalence of MTHFR mutation in healthy individuals in Erbil northern Iraq. Determining such prevalence rates and comparing the data with those found in other regions.

METHODS

The total studied sample consisted of 100 apparently healthy individuals attending the premarital screening center in Erbil for routine mandatory checkup and included 56 male and 44 female and with age ranges of 16-45 and 15-38 years respectively. From each individual 3 mL of whole blood were collected in EDTA for isolation of genomic DNA using a phenol chloroform method.¹¹

The C677T *MTHFR* gene mutation was detected by polymerase chain reaction and restriction fragment length polymorphism (PCR-RFLP) analysis using *Hinf* I restriction enzyme according to Frosst et al.⁸ The PCR reactions were done in a sterile 0.2 ml tube, the reaction mix was of a final volume of 25 µL containing 1×PCR buffer (Promega – USA) with 1.5 mmol/L MgCl₂, 1.5 unit Taq DNA polymerase, 10 µmol/L dNTP, DNA template 2.5 µl, primers (5'-TGAAGGAGAAGGTGTCTGCGGGA-3' and 5'-AGGACGGTGCGGTGAGAGTG-3') (0.5µl each), completed to 25 µl with sterile distilled water. PCR Conditions for *MTHFR* 677: Pre-PCR: 94°C for 8 Min, then 40 cycles of: denaturation:94°C for 1 min, annealing:63°C for 1 min, extension :72°C for 1 min, followed by final extension for 7 min.¹² PCR products were

checked on 2% of agarose gels followed by staining with ethidium bromide to stain DNA fragments. The amplified PCR products (*MTHFR*) were subjected to *Hinf* I restriction enzyme digestion at 37°C overnight. Digestion was carried out in a final volume of 10µL, using 8.5µL of PCR product, 5 units of *Hinf*I enzyme, and 1.0µL of buffer. Size analysis of the restriction fragments were visualized by gel electrophoreses of digested PCR products using 3% agarose and stained with ethidium bromide. Polymorphism C677T creates a recognition sequence for the restriction enzyme *Hinf*I, and this is detected by digestion of the 198-bp PCR product, generating 23- and 175-bp fragments for the polymorphism in homozygous state (genotype TT). Genotype CC was characterized by the presence of a 198-bp fragment, and genotype CT was characterized by the presence of three fragments, 198 bp, 175 bp and 23 bp.¹³

RESULTS

Amplification using specific primers on DNA extracted on the 100 apparently healthy individuals yielded a 198 bp product. After restriction digestion of PCR products with *Hinf*I: 6 samples (6%) had full digestion yielding 175 bp (the 23 bp was too faint to be detected on agarose gel) and thus were labeled as homozygous (TT genotype), 37 (37%) had the 175 in addition to the 198 bp fragments, thus labeled as heterozygous (CT genotype), while the remaining 57 Ampicons (57%) did not show any digestion and just retained the original 198 kb fragment, and thus labeled as Wild genotype (CC) (Table 1) and (Figure 1).

Table 1. The number and frequency MTHFR 677CT genotype mutations found in healthy individuals from Erbil –Iraq (100 healthy individuals)

Sex of individuals	Homozygous (TT) No. (%)	Heterozygous (CT) No. (%)	Wild (CC) No. (%)
Males (n= 56)	4 (7)	23(41)	29 (51.8)
Females (n= 44)	2 (4.5)	14(31.8)	28 (63.6)
Total	6(6)	37(37)	57(57)

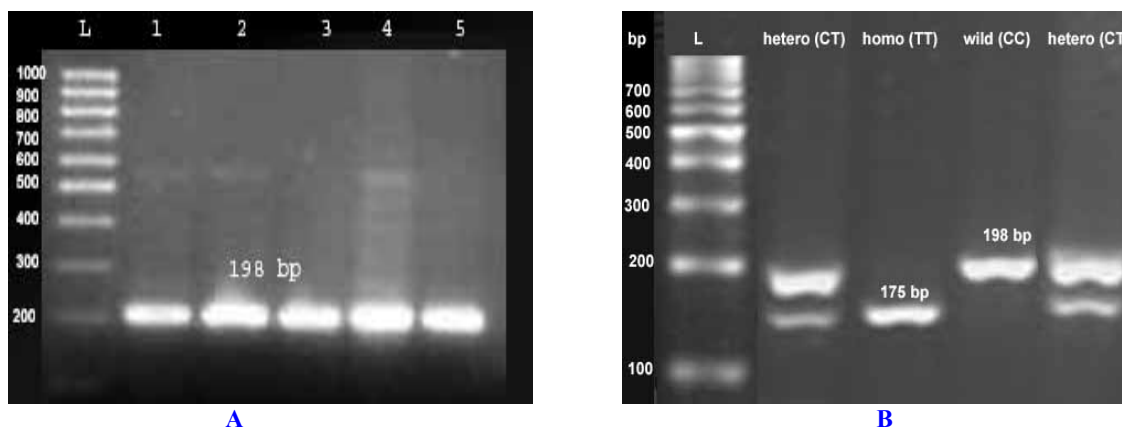


Figure 1. A. Represent the PCR product MTHFR C677T genotype bands in 198bp fragments. Polymorphism analysis of the methylenetetrahydrofolate reductase gene (MTHFR) amplicons by agarose gel electrophoresis after restriction endonuclease digestion (RFLP). CT genotype yields 198bp and 175bp TT genotype (175bp), and CC (198bp). L: Ladder molecular weight.

DISCUSSION

Erbil (Hewlêr in Roman-alphabet Kurdish) (also written Irbil) is a large city in northern Iraq with a population of approximately 1.3 million (2009), bordering Turkey to the north and Iran to the east. It is largely populated by Kurds but has a small minority of Assyrians.

This study showed that the frequency of homozygosity to MTHFR C677T mutation was 6% which is lower than that reported in previous study from Duhok to the north of Erbil, where a frequency of 8% was found.¹⁴ Table 2 demonstrates that the frequency of homozygosity to MTHFR C677T mutation was 6% also lower than those reported from neighboring Syria,¹⁵ Turkey,¹⁶ Jordan,¹⁷ Tunisia,¹⁸ some European countries,¹⁹ Brazil,¹³ USA,^{20,21} Japan,²² china,²³ and Korea.²⁴

However it was higher than that reported from Iran,²⁵ Oman,²⁶ Bahrain,²⁷ Egypt,²⁸ Northern Italy,²⁹ Pakistan,³⁰ and North India.³¹ However and in contrast to the above reports, homozygosity for this polymorphism is almost absent in Africans and people of African descent.^{21,22}

A study confirmed a relatively high frequency of the 677TT genotype in the French population and an association with elevated tHcy concentration in men but not in women.³² Individuals homozygous for

the C677T mutation have moderately increased concentrations of fasting plasma homocysteine especially in the presence of low (<15.4 mol/L) plasma folate.³³ The current study did not however include studying homocysteine levels in those with the mutation, which may have added valuable information.

Several previous studies revealed that a very common mutation in the MTHFR gene C677T is related to mild homocysteinemia and might increase the risk for vascular occlusive pathology. However, other recent publications negate this relationship.^{34,35,36} In their important study, Frosst and colleagues not only identified the MTHFR mutation responsible for thermolability of the enzyme but also established that subjects homozygous for this mutation had elevated fasting and postmethionine plasma homocyst(e)ine concentrations.⁸ The association between the mutation and elevated fasting homocyst(e)ine was confirmed also by van der Put and colleagues in 1995, who found that the mutation was associated with decreased MTHFR activity.³⁷ Homozygosity for the mutation, and to a lesser extent heterozygosity, were associated with moderately increased fasting tHcy levels.³⁸⁻⁴⁰

Table 2. The frequencies of the MTHFR (C677T) Homozygous (TT) % mutation in Eastern Mediterranean and worldwide studies

Location	MTHFR (C677T) Homozygous (TT) %	References
Iraq/ Erbil	6	Current study
Iraq/ Duhok	8	14
Turkey	9.6	16
Iran	5	25
Syria	18	15
Jordan	8	17
Oman	2.45	26
Bahrain	2.6	27
Egypt	5	28
Tunisia	7	18
Europe	8-18	19
USA	13	20,21
Brazil	9.6	13
Africa/African origin	0	21,22
Japan	11.5	22
Pakistan	1	30
China	17.8	23
North India	1.5	31
Northern Italy	4	29
Korean	14.05	24

The study of Peadar, and coworker revealed that heterozygosity for the MTHFR polymorphism, which is present in 38% of the population, increases the risk of neural tube defects. Most studies of MTHFR C677T and neural tube defects and other conditions have focused on the risk associated with T allele homozygosity. The possibility that heterozygosity might also increase neural tube defect risk has gone unrecognised except for a small study in which an association between CT and these malformations was thought to be due to the higher than expected proportion of CC control subjects.⁴¹ The individuals with MTHFR C677T C/T and T/T genotypes were at a higher risk of developing colon cancer, but they were at a decreased risk of developing rectal cancer.^{42,23} Polymorphism at C677T shows marked heterogeneity based upon ethnicity and geographical location.⁴³ Only individuals who were homozygous (TT) for the

C677T mutation had significantly higher plasma total homocysteine concentrations, which is in accordance with what has been reported previously.^{8,38} The data demonstrate that 677C T polymorphisms, whether homozygous or heterozygous, are significantly associated with anemia sickle disease (ASD). The homozygous (TT) individuals are reported to have an approximately 50% decrease in MTHFR enzyme activity, and the heterozygous (CT) a 30% decrease in enzyme activity as measured in their lymphocytes.²¹

Finally methylenetetrahydrofolate reductase C677T mutation is commonly encountered among healthy individuals in Erbil city, although it was rather less frequent than that documented earlier in Duhok, to the north of the former city. The clinical implications of our finding require further clinical studies.

REFERENCES

1. Oliveira KC, Bianco B, Verreschi IT, Guedes AD, Galera BB, Galera MF, et al. Prevalence of the polymorphism MTHFR A1298C and not MTHFR C677T is related to chromosomal aneuploidy in Brazilian Turner Syndrome patients. *Arq Bras Endocrinol Metabol.* 2008;52(8):1374-81.
2. Ueland PM, Hustad S, Schneede J, Refsum H, Vollset SE. Biological and clinical implications of the MTHFR C677T polymorphism. *Trends Pharmacol Sci.* 2001;22(4):195-201.
3. Friso S, Choi SW, Girelli D, Mason JB, Dolnikowski GG, Bagley PJ, et al. A common mutation in the 5,10-methylenetetrahydrofolate reductase gene affects genomic DNA methylation through an interaction with folate status. *Proc Natl Acad Sci U S A.* 2002;99(8):5606-11. Epub 2002 Apr 2.
4. Föding M, Hörl WH, Sunder-Plassmann G. Molecular biology of 5,10-methylenetetrahydrofolate

- reductase. *J Nephrol.* 2000;13(1):20-33.
5. Schneider JA, Rees DC, Liu YT, Clegg JB. Worldwide distribution of a common methylenetetrahydrofolate reductase mutation. *Am J Hum Genet.* 1998;62(5):1258-60.
6. Hultberg B, Agardh E, Andersson A, Brattström L, Isaksson A, Israelsson B, et al. Increased levels of plasma homocysteine are associated with nephropathy, but not severe retinopathy in type 1 diabetes mellitus. *Scand J Clin Lab Invest.* 1991;51(3):277-82.
7. Kark JD, Selhub J, Adler B, Gofin J, Abramson JH, Friedman G, et al. Nonfasting plasma total homocysteine level and mortality in middle-aged and elderly men and women in Jerusalem. *Ann Intern Med.* 1999;131(5):321-30.
8. Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, et al. A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. *Nat Genet.* 1995;10(1):111-3.
9. Mayer EL, Jacobsen DW, Robinson K. Homocysteine and coronary atherosclerosis. *J Am Coll Cardiol.* 1996;27(3):517-27.
10. Stampfer MJ, Malinow MR, Willett WC, Newcomer LM, Upson B, Ullmann D, et al. A prospective study of plasma homocyst(e)ine and risk of myocardial infarction in US physicians. *JAMA.* 1992;268(7):877-81.
11. Ausubel FM, Brent R, Kingston RE, Moore DD, Seidman JG, Smith JA, et al. Current protocols in molecular biology. 2nd ed. New York: Wiley Interscience; 1987.
12. Deeparani T, Pillai MR, Elavazhagan T. Detection of MTHFR C677T and A1298C gene polymorphism in congenital heart disease. *Middle-East Journal of Scientific Research.* 2009;4(2):127-32.
13. Oliveira KC, Bianco B, Verreschi IT, Guedes AD, Galera BB, Galera MF, et al. Prevalence of the polymorphism MTHFR A1298C and not MTHFR C677T is related to chromosomal aneuploidy in Brazilian Turner Syndrome patients. *Arq Bras Endocrinol Metabol.* 2008;52(8):1374-81.
14. Al-Allawi NA, Jubrael JM, Baban NK, BABAN NK, Gedeon GS. Thrombophilic Mutations in Blood Donors in Duhok / Iraq. *Duhok Med J.* 2009;3(1):25-32.
15. Herrmann W, Obeid R, Jouma M. Hyperhomocysteinemia and vitamin B-12 deficiency are more striking in Syrians than in Germans--causes and implications. *Atherosclerosis.* 2003;166(1):143-50.
16. Sazci A, Ergul E, Kaya G, Kara I. Genotype and allele frequencies of the polymorphic methylenetetrahydrofolate reductase gene in Turkey. *Cell Biochem Funct.* 2005;23(1):51-4.
17. Eid SS, Rihani G. Prevalence of factor V Leiden, prothrombin G20210A, and MTHFR C677T mutations in 200 healthy Jordanians. *Clin Lab Sci.* 2004;17(4):200-2.
18. Mtiraoui N, Zammiti W, Ghazouani L, Braham NJ, Saidi S, Finan RR, et al. Methylenetetrahydrofolate reductase C677T and A1298C polymorphism and changes in homocysteine concentrations in women with idiopathic recurrent pregnancy losses. *Reproduction.* 2006;131(2):395-401.
19. Gemmati D, Serino ML, Trivellato C, Fiorini S, Scapoli GL. C677T substitution in the methylenetetrahydrofolate reductase gene as a risk factor for venous thrombosis and arterial disease in selected patients. *Haematologica.* 1999;84(9):824-8.
20. Folsom AR, Cushman M, Tsai MY, Aleksic N, Heckbert SR, Boland LL, et al. A prospective study of venous

- thromboembolism in relation to factor V Leiden and related factors. *Blood*. 2002;99(8):2720-5.
21. Boris M, Goldblatt A, Galanko J, James SJ. Association of MTHFR gene variants with autism. *Journal of American Physicians and Surgeons*. 2004;9(4):106-8.
22. Botto LD, Yang Q. 5,10-Methylenetetrahydrofolate reductase gene variants and congenital anomalies: a HuGE review. *Am J Epidemiol*. 2000;151(9):862-77.
23. Cao HX, Gao CM, Takezaki T, Wu JZ, Ding JH, Liu YT, et al. Genetic polymorphisms of methylenetetrahydrofolate reductase and susceptibility to colorectal cancer. *Asian Pac J Cancer Prev*. 2008;9(2):203-8.
24. Oh D, Kim NK, Jang MJ, Kim HC, Lee JH, Lee JA, et al. Association of the 5,10-methylenetetrahydrofolate reductase (MTHFR C677T and A1298C) polymorphisms in Korean patients with adult acute lymphoblastic leukemia. *Anticancer Res*. 2007;27(5A):3419-24.
25. Golbahar J, Fathi Z, Tamadon M. Distribution of 5,10-methylenetetrahydrofolate reductase (C667T) polymorphism and its association with red blood cell 5-methyltetrahydrofolate in the healthy Iranians. *Clin Nutr*. 2005;24(1):83-7.
26. Pathare A, Al Kindi S, Al Haddabi H, Dennison D, Bayoumi R, Muralitharan S. Hereditary thrombophilia in ethnic Omani patients. *Am J Hematol*. 2006;81(2):101-6.
27. Al-Habboubi H, Tamim H, Ameen G, Almawi WY. C677T and A1298C single nucleotide polymorphisms in the methylenetetrahydrofolate reductase gene among Bahraini Arabs. *Thromb Haemost*. 2004;91(4):843-5.
28. Mackawy AM, Badawy ME. Methylenetetrahydrofolate reductase gene polymorphism and the risk of ischemic stroke in type 2 diabetic Egyptian patients. *Global Journal of Health Science*. 2011;3(2):162-74.
29. Ferrazzi P, Di Micco P, Quaglia I, Rossi LS, Bellatorre AG, Gaspari G, et al. Homocysteine, MTHFR C677T gene polymorphism, folic acid and vitamin B 12 in patients with retinal vein occlusion. *Thromb J*. 2005;7;3:13.
30. Michael S, Qamar R, Akhtar F, Khan WA, Ahmed A. C677T polymorphism in the methylenetetrahydrofolate reductase gene is associated with primary closed angle glaucoma. *Mol Vis*. 2008;14:661-5.
31. Tripathi R, Tewari S, Singh PK, Agarwal S. Association of homocysteine and methylenetetrahydrofolate reductase (MTHFR C677T) gene polymorphism with coronary artery disease (CAD) in the population of North India. *Genet Mol Biol*. 2010;33(2):224-8. Epub 2010 Jun 1.
32. Chango A, Boisson F, Barbé F, Quilliot D, Drosch S, Pfister M, et al. The effect of 677C-->T and 1298A-->C mutations on plasma homocysteine and 5,10-methylenetetrahydrofolate reductase activity in healthy subjects. *Br J Nutr*. 2000;83(6):593-6.
33. Cedergren MI, Selbing AJ, Källén BA. Risk factors for cardiovascular malformation--a study based on prospectively collected data. *Scand J Work Environ Health*. 2002;28(1):12-7.
34. Verhoeff BJ, Trip MD, Prins MH, Kastelein JJ, Reitsma PH. The effect of a common methylenetetrahydrofolate reductase mutation on levels of homocysteine, folate, vitamin B12 and on the risk of premature atherosclerosis. *Atherosclerosis*. 1998;141(1):161-6.
35. Abbate R, Sardi I, Pepe G, Marcucci R, Brunelli T, Prisco D, et al. The high prevalence of thermolabile 5-10 methylenetetrahydrofolate reductase (MTHFR) in Italians is not associated to an increased risk for coronary artery

- disease (CAD). *Thromb Haemost.* 1998;79(4):727-30.
36. Al-Allawi NA, Avo AS, Jubrael JM. Methylenetetrahydrofolate reductase C677T polymorphism in Iraqi patients with ischemic stroke. *Neurol India.* 2009;57(5):631-5.
37. van der Put NM, Steegers-Theunissen RP, Frosst P, Trijbels FJ, Eskes TK, van den Heuvel LP, et al. Mutated methylenetetrahydrofolate reductase as a risk factor for spina bifida. *Lancet.* 1995;346(8982):1070-1.
38. Jacques PF, Bostom AG, Williams RR, Ellison RC, Eckfeldt JH, Rosenberg IH, et al. Relation between folate status, a common mutation in methylenetetrahydrofolate reductase, and plasma homocysteine concentrations. *Circulation.* 1996;93(1):7-9.
39. Christensen B, Frosst P, Lussier-Cacan S, Selhub J, Goyette P, Rosenblatt DS, et al. Correlation of a common mutation in the methylenetetrahydrofolate reductase gene with plasma homocysteine in patients with premature coronary artery disease. *Arterioscler Thromb Vasc Biol.* 1997;17(3):569-73.
40. Clarke R, Woodhouse P, Ulvik A, Frost C, Sherliker P, Refsum H, et al. Variability and determinants of total homocysteine concentrations in plasma in an elderly population. *Clin Chem.* 1998;44(1):102-7.
41. Kirke PN, Mills JL, Molloy AM, Brody LC, O'Leary VB, Daly L, et al. Impact of the MTHFR C677T polymorphism on risk of neural tube defects: case-control study. *BMJ.* 2004;328(7455):1535-6. Epub 2004 May 21.
42. Kim DH, Ahn YO, Lee BH, Tsuji E, Kiyohara C, Kono S. Methylenetetrahydrofolate reductase polymorphism, alcohol intake, and risks of colon and rectal cancers in Korea. *Cancer Lett.* 2004;216(2):199-205.
43. Mansoor A, Mazhar K, Ali L, Muazzam AG, Siddiqi S, Usman S. Prevalence of the C677T single-nucleotide polymorphism in the methylenetetrahydrofolate reductase gene among Pakistani ethnic groups. *Genet Test Mol Biomarkers.* 2009;13(4):521-6.

پوخته

گهورینا جینی C677T تیتراهایدروفولیت ریدەکتیز MTHFR دناڤ کەسین ساخەمدا ل هەولێری – عێراق

پێشەکی و ئارمانج: ئەنزیمی تیتراهایدروفولیت ریدەکتیز MTHFR ئێکە ژ ئەنزیمن سەرەکی یێن ریکخەر بو میتابولیزما هوموسستینی. ئێک ژ گهورینن مشە د جینی قی ئەنزیمی دا گهورینا C→T677 کو دبینە ئەگەری کیمبوونا کاری ئەنزیمی و بلندبوونا ریزا هوموسستینی د خوینی دا. هەڤبەندی دناڤهرا قی گهورینی دا و نهخوشیپن دەمارا باش یا دیار نینه، و دبیت کارلێکرن هەبیت ژ بنەمای نژادی ڤه. ئارمانجا قی ڤهکولینی دیارکرن مشەتیا قی گهورینا جینی دناڤ کەسین ساخەمدا ل هەولێری – عێراق.

ریکین ڤهکولینی: سەرجهمی 100 کەسین ساخەم یێن سەرەدانا سەنتەری پشکنینن بەری مارەبرینی ل هەولێری کری هاتنه بەشداریکرن ڤهکولینی دا. فرەشیوہیا جینی C677T (MTHFR) هاته پشکنینکرن بو هەمی بەشداریبويا بریکا PCR و RFLP بو پارچا DNA ل نافکوکا 677 بو هەمیان.

ئەنجام: گهورینا C677T MTHFR هاته دیتن بشیوی ئێک جور ل 6% و هەمەجور ل 37% ژ بەشداریبويا.

دەرئەنجام: گهورینا C677T MTHFR مشە دەیتە دیتن ل دەف کەسین ساخەمدا ل هەولێری، هەرچەندە ریزە کیمترە ژ یا بەری نوکە هاتیه دیتن ل دەوکی لباکوری هەولێری. کارتیکنن کلینیکی یێن قی ڤهکولینی پێدقی ب پتر ڤهکولینایه.

الخلاصة

الطفرة في جين C677T (Methylenetetrahydrofolate reductase MTHFR) في

الاصحاء في مدينة اربيل - العراق

خلفية واهداف البحث: يعد انزيم (methylenetetrahydrofolate reductase (MTHFR احد الانزيمات الرئيسية المنظمة لايض ال homocysteine . ان الطفرة A 677C→T في جين MTHFR ينتج انخفاض في الفعالية الانزيمية و هذا يساهم في زيادة homocysteine في البلازما . العلاقة بين الطفرة C677T في جين ال MTHFR المرض الوعائي لاتزال غير مؤكدة وربما تتأثر بالعرق. نهدف الدراسة الحالية الى احتساب تردد الطفرة C677T في انخفاض الاصحاء في مدينة اربيل / عراق.

طرق البحث: جمعت عينات من 100 شخص اصحاء متطوعون من المراجعين الى مركز الفحص قبل الزواج في مدينة اربيل-العراق. تم تقدير تعدد اشكال الجين الطافر MTHFR C677T في كل العينات باستعمال تقنيات ال PCR-RFLP.

النتائج: وجد أن نسبة الجين الطافر MTHFR C677T في متماثل الزيجة 6% بينما في متباين الزيجة 37%.
الاستنتاجات: اتضح ان تردد الطفرة في جين MTHFR C677T شائعا في الاشخاص الاصحاء في مدينة اربيل وبالاخرى كان هذا التردد اقل من تردد نفس الجين في دراسات مبكرة في مدينة دهوك شمال مدينة اربيل. ان النتائج السريية الحالية تحتاج الى دراسات سريرية اكثر.

CHILDHOOD HENOC-SCHÖNLEIN PURPURA IN KURDISH POPULATION OF DUHOK CITY

KHALID N. ABDURRAHMAN, MBChB, DCH, FIBMS*
DELAWER B. MIKAEL, MBChB, DCH**

Submitted 24 Mar 2012; accepted 5 Jun 2012

ABSTRACT

Background and objectives Henoch-Schönlein purpura is the most common systemic vasculitis in children, mainly affecting the skin, joints, gastrointestinal tract, and kidneys. This study was designed to assess the clinical and epidemiological characteristics of Kurdish children with Henoch-Schönlein purpura in Duhok city.

Methods A prospective study was conducted on children diagnosed as Henoch-Schönlein purpura, who were treated in Heevi pediatric teaching hospital between July, 2009 and July, 2011. The collected data included age, sex, the initial presenting feature, associated systemic clinical features, season of presentation, and associated triggering factors. Laboratory investigations included complete blood count, erythrocyte sedimentation rate, renal function tests and urinalysis. Skin biopsy was taken from 11 patients. Patients were followed up by urinalysis and renal function tests for 6 months to pick up renal involvement.

Results Over the 2 years period, a total of 51 patients were diagnosed with Henoch-Schönlein purpura, of whom 28 were males and 23 females. The patients were aged between 10 months-15 years, the mean age at presentation was 7.1 years, with a male to female ratio of 1.2:1. Disease onset was more common in winter and autumn. All cases had palpable purpura. Large joint arthritis/arthralgia occurred in 41 (80.4%), abdominal pain in 24 (47%) patients, and renal involvement in one patient. The result of skin biopsy was consistent with the diagnosis of Henoch-Schönlein purpura. Complete recovery occurred in all patients. After 6 months of follow-up, no patient had renal involvement.

Conclusions Henoch-Schönlein purpura in Kurdish children is milder with fewer renal manifestations than that in the previously published studies in other areas.

Duhok Med J 2012;6(1): 43-53.

Key words: Henoch-Schönlein purpura, Purpura, Arthritis, Skin biopsy

Kawasaki disease, most practicing pediatricians will never encounter a case.¹ HSP is a systemic small vessel vasculitis characterized by non-thrombocytopenic palpable purpura, arthritis, bowel angina and hematuria/proteinuria.²⁻⁴ Although HSP is a condition that can occur from age 6 months to adulthood, 50% of cases occur in children under 5 years of age and 75% are under 10 years. In most reports HSP is more common in boys.⁵ This disease is usually self-limiting. Involvement of

internal organs such as kidney, intestine and central nervous system are the major complications.²⁻⁴ The prognosis is thought to be good as long as the patients have no renal symptoms.⁶

Although HSP is not uncommon in children, there are few large-scale epidemiological studies of childhood HSP, especially nationwide surveys.⁷⁻⁹ There is no existing data in Kurdistan region (North of Iraq) on this topic.

In this study, we prospectively evaluated the epidemiological and clinical

*Assistant professor of Pediatrics, Department of Pediatrics, School of Medicine, Faculty of Medical Sciences, University of Duhok - Heevi Pediatric Teaching Hospital

**Heevi Pediatric Teaching Hospital

Correspondence author: Khalid N. Abdurrahman. Email: drkhnawaf@yahoo.com

data, main laboratory abnormalities and outcome in 51 Kurdish children with HSP followed by a single center, over a follow-up period of 6 months.

METHODS

All Kurdish children with HSP who were diagnosed by pediatricians and treated in Heevi Hospital in Duhok city, between July 2009 and July 2011 included in this study (Non-Kurdish children were excluded). The diagnosis was made based on the American College of Rheumatology criteria.¹⁰ Patients were diagnosed as HSP if they had palpable purpura not related to thrombocytopenia, with or without other manifestations. An atypical HSP case was defined as a HSP patient without skin rash in the first 24 hours of admittance.¹¹ The demographic (age and sex), and clinical (recent history of febrile illness, seasons of occurrence, presence of purpura, joint involvement, gastrointestinal manifestations, renal involvement, other organ involvements) characteristics were collected and analyzed.

Laboratory investigations included complete blood count (CBC) and erythrocyte sedimentation rate (ESR), blood urea and serum creatinine. In addition, repeated urinalyses were undertaken to detect renal involvement.

HSP nephritis (HSN) was defined as the presence of gross or microscopic hematuria with or without proteinuria.¹² The following definitions were adopted for renal involvement: microscopic hematuria (> 10 erythrocytes/HPF), macroscopic hematuria (> 100 erythrocytes/HPF), proteinuria (> 5 mg/kg/24 h).¹² Ultrasound of the abdomen was done in all patients with abdominal pain.

Skin biopsy from the involved skin was taken from 11 patients and subjected to light microscopy. The patients were followed up every 2 weeks for the first 3 months and every month for the next 3 months by urinalysis and renal function tests. This study received prior approval from the Medical Ethics Committee at the College of Medicine, University of Duhok.

RESULTS

Over the 2 years period, a total of 51 patients were diagnosed with HSP, of whom 28 (54.9%) were males and 23 (45.1%) females and the male-to-female ratio was 1.2:1. The age at presentation ranged between 10 months to 15 years old, and the mean age at presentation was 7.1 years. Figure 3.1 shows the distribution of number of cases according to patients' age and sex. Most cases presented between 5 and 10 years ($n=45$; 88.2% of total cases).

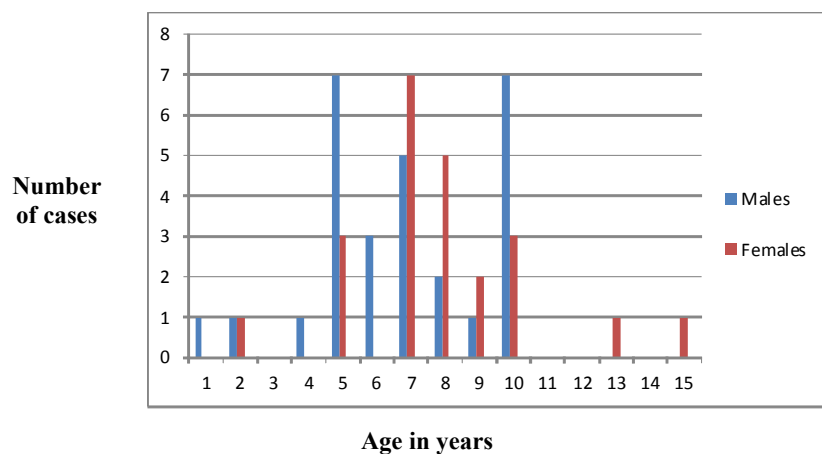


Figure 1. Age and sex distribution of patients

Figure 3.2 shows that HSP could occur year-round, but more commonly in winter and autumn than in other seasons. The clinical data of patients are shown on table 3.1.

Thirty-three (64.7%) patients had history of respiratory infection about 1-3

weeks before the onset of the disease. The presenting symptoms were skin rash (Figure 3.3) in 51 (100%), joint symptoms in 41 (80.4%) and abdominal pain in 24 (47%) patients. The mean duration of symptoms before diagnosis was 2 days (range 1-4 days).

■ Winter(22) ■ Spring(9) ■ Summer(5) ■ Autumn(15)

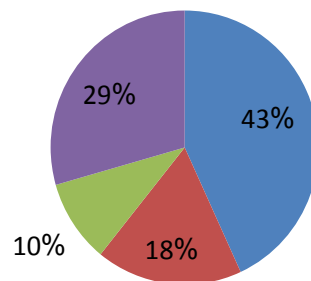


Figure 3.2 Seasonal variations in Henoch-Schönlein purpura cases

Table 3.1. Clinical data of patients (n=51)

Clinical data	No. (%)
History of respiratory infection	33 (64.7)
Purpura	51 (100)
Lower limb	51 (100)
Buttocks	29 (56.8)
Upper limbs	21 (41.1)
Face	1 (1.96)
Arthritis/Arthralgia	41 (80.4)
Abdominal pain	24 (47)
Melena	1 (1.96)
Edema of extremities	13 (25.5)
Microscopic hematuria	1 (1.96)
Fever	8 (15.6)
First manifestation	
Purpura	42 (82.3)
Arthritis/Arthralgia	5 (9.8)
Abdominal pain	4 (7.8)



Figure 3.3. The classic rash of Henoch-Schönlein purpura

The variability of cutaneous morphology was noted in one patient. This variation was found in a 9 year old girl who presented with hemorrhagic vesicles and bullae varying in size from 10 to 30 mm in diameter, on both legs, and dorsum of feet, in addition to palpable purpuric rash (Figure 3.4).



Figure 3.4. Hemorrhagic vesicles, a rare presentation of Henoch-Schönlein purpura. Skin biopsy demonstrated leukocytoclastic vasculitis

One patient (1.96 %) was infant (10 months old boy), and was admitted for a purpuric rash observed 1 day earlier. At the time of admission he was febrile and had a purpuric rash on his face, pinnae, and extremities. His hands and legs were edematous (Figure 3.5 A, B, and C).



A



B



C

Figure 3.5 A, B, and C. An infant with Henoch-Schönlein purpura on face, ears, and extremities. Skin biopsy demonstrated leukocytoclastic vasculitis

Only one patient (1.96%) had microscopic hematuria without proteinuria and normalization of urinary abnormalities occurred over a period of 2 weeks. No cases had central nervous system or pulmonary manifestations.

Elevated ESR (>30 mm/h) was observed in 16 (31.3%) patients, and mild thrombocytosis ($450-500 \times 10^9$ /L) in 19 (37.2%) patients. Other blood elements were normal. Renal function tests were normal, even in the patient with hematuria.

Ultrasound abdomen was normal in all patients with abdominal pain.

The skin biopsy was performed in 11 patients (one was an infant, one had hemorrhagic vesicles, 5 had skin and joint manifestations, and 4 had skin and GIT manifestations). Light microscopy of skin biopsy showed perivascular infiltrate of polymorphonuclear leukocytes, nuclear dust, extravascular erythrocytes, and fibrinoid necrosis of the vessel walls. These findings are consistent with leukocytoclastic vasculitis. Direct immunofluorescence examination to reveal the presence of IgA and C3 in the mesangium and capillary wall was not done (not available).

The average duration of the disease was 9 days (range 6-14 days) and all patients recovered uneventfully. None of the patients who escaped renal involvement at initial presentation have developed any urinary abnormalities on follow-up. Only 3 patients were lost to follow up after one month.

DISCUSSION

Although few cases of HSP might have been diagnosed and treated elsewhere (e.g. dermatology or private clinics), we have seen 51 cases of HSP over a period of 2 years. To our knowledge, the present study is the first one from Iraq where the clinical diagnosis has been confirmed by histopathological study on skin biopsy in some of the patients. The diagnosis of HSP in this series was made according to the

American College of Rheumatology criteria,¹⁰ though a new and more realistic criteria, EULAR/PreS endorsed consensus criteria for classification of childhood vasculitides,¹³ has been proposed. We believe that both methods can be applied to our patients with the same results, because all patients had joint and/ or GIT manifestations in addition to skin involvement, and no patient showed isolated palpable purpura (i.e., without other features).

In this study, the mean age of patients at presentation (7.1 years) and the male-to-female ratio (1.2:1) were comparable to that reported from Turkey,¹¹ Jordan,¹⁴ India,¹⁵ and Taiwan,¹⁶ while some studies have shown a female predominance.^{17,18} These variations may be attributable to the small sample sizes in the majority of previous studies and different time frames, races and geographical areas from which the data were recorded and analyzed. The great majority of our patients (72%) presented during winter and autumn, and 64.7% of patients had a respiratory infection prior to onset of the disease. A similar seasonal pattern was noticed by other authors.^{11,14,16} The disease clustering in winter and the histories of preceding upper respiratory infections recorded in many HSP patients,^{4,11,19,20} provide clues to the possibility that HSP is infection-related, but drugs (antibiotics, ACE inhibitors, NSAIDs) and certain toxins (insect bites, vaccinations and food allergies) have also been implicated.^{18,21} These epidemiological characteristics may be valuable for disease prevention and for the further etiological studies of HSP.

All patients in this study had skin involvement (purpura), which appeared as the first manifestation at onset of disease in 82.3%, while abdominal pain and arthritis preceded the rash in 4 (7.8%) and 5 (9.8%) patients, respectively. This is in contrast to the results reported by Kumar *et al.*¹⁵ where the rash appeared as the first manifestation in 47% and abdominal pain preceded the rash in 29% patients.

Occurrence of severe acute abdominal pain in the absence of skin rash may lead to misdiagnosis of acute abdomen and unnecessary surgical exploration.^{21,22}

One patient was found to have hemorrhagic bullae and vesicles, in addition to purpuric rash. Bullous eruption in HSP, as in our case, appears often in adults while it is very rare in children.²³ Clinicians must be aware that HSP can present with atypical features that make the diagnosis difficult.

Transient arthritis/arthralgia involving large joints and GIT involvement (abdominal pain) were comparable to that reported in other studies,^{18,19,21,24,25} though serious GIT complications (e.g., hemorrhage, perforation and intussusceptions) were absent.

The youngest child among our patients was 10 months old boy, who was the only patient under 2 years of age in our sample. The occurrence of HSP in infants is considered rare.^{21,26} However, infantile hemorrhagic edema has been described, and the question is asked whether this is a very early form of HSP or a different vasculitis altogether.^{27,28}

Renal involvement (nephritis) is potentially the most worrisome feature of HSP, and the majority of these (85%) doing so within the first 4 weeks and 97% within six months.²⁹ Only one of our patients (1.96%) had microscopic hematuria during the acute stage of the disease, in contrast to the higher incidence reported by other authors (13.6-45%).^{11,14,15,21,30}

After 6 months of follow-up, no patient had renal involvement. There is no need to follow up after the first six months those whose urinalysis remains normal, but measurements of serum urea and creatinine need to continue in the presence of continued urinary abnormalities.²⁹

According to other studies in the medical literature,³¹⁻³⁴ renal involvement is the principal determinant of severity and prognosis in HSP, therefore, the lower rate of renal involvement in our patients is an

indicator of mild pattern of the disease and it suggests ethnic differences for nephritis in HSP, which was reported by other authors.³⁵

In the pediatric population, skin biopsy is reserved for patients with an unusual presentation of HSP (no rash or atypical rash) and patients with significant renal disease.³⁶ However, skin biopsies have been suggested as possible diagnostic criteria for HSP, and are commonly used as such in the case of adults.³⁷ Owing to the typical presentation of our patients, we performed skin biopsy in 11 cases. The light microscopy with hematoxylin and eosin stains demonstrated a classic leukocytoclastic vasculitis in postcapillary venules in the 11 patients.

Elevated ESR and thrombocytosis are well documented features of HSP,^{18,38} and were seen in 31.7% and 46% of our patients, respectively. Thrombocytosis helps in distinguishing this form of purpura from that caused by thrombocytopenia.³⁸ The degree of thrombocytosis is believed to correlate with severity of illness,^{5,39} though this could not be documented in our patients.

The frequency of relapses varies from series to series. Relapses occurred in 33% of American patients,⁷ in 5% of Turkish patients,¹¹ in 15% of Spanish patients,¹⁸ and in 35% of Italian patients.⁴⁰ In our study group, all patients recovered uneventfully and neither chronicity nor relapse was recorded during the period of follow up (6 months). However, patients with HSP should be followed over the long term, since recurrence is likely to occur, especially during a 2-year period after the first outbreak.^{4,25} The absence of relapse of HSP in our patients may be explained, to some extent, by mild pattern of the disease and absence of renal involvement which are regarded as possible predictive factors for relapse.^{33,34}

In conclusion, the demographic and epidemiological characteristics of HSP in Kurdish children are comparable to that reported from other countries, but it is

milder with lower rate of renal involvement. It appears that ethnic differences play a role in the presentation pattern of HSP. Larger studies are required to confirm the results in this study, to monitor the long-term outcome of the disease and to compare with other ethnic groups in the same area.

REFERENCES

1. Dedeoglu F, Sundel RP. Vasculitis in children. *Rheum Dis Clin North Am*. 2007;33(3):555-83.
2. Bissonnette R, Dansereau A, D'Amico P, Pateneau J, Paradis J. Perforation of large and small bowel in Henoch-Schönlein purpura. *Int J Dermatol*. 1997;36(5):361-3.
3. Kawasaki Y, Suzuki J, Nozawa R, Suzuki S, Suzuki H. Efficacy of methylprednisolone and urokinase pulse therapy for severe Henoch-Schönlein nephritis. *Pediatrics*. 2003;111(4 Pt 1):785-9.
4. Saulsbury FT. Henoch-Schönlein purpura. *Curr Opin Rheumatol*. 2001;13(1):35-40.
5. Tizard EJ. Henoch-Schönlein purpura. *Arch Dis Child*. 1999;80(4):380-3.
6. Niaudet P, Murcia I, Beaufils H, Broyer M, Habib R. Primary IgA nephropathies in children: prognosis and treatment. *Adv Nephrol Necker Hosp*. 1993;22:121-40.
7. Saulsbury FT. Henoch-Schönlein purpura in children. Report of 100 patients and review of the literature. *Medicine (Baltimore)*. 1999;78(6):395-409.
8. Stewart M, Savage JM, Bell B, McCord B. Long term renal prognosis of Henoch-Schönlein purpura in an unselected childhood population. *Eur J Pediatr*. 1988;147(2):113-5.
9. Watts RA, Jolliffe VA, Grattan CE, Elliott J, Lockwood M, Scott DG. Cutaneous vasculitis in a defined population--clinical and epidemiological associations. *J Rheumatol*. 1998;25(5):920-4.
10. Mills JA, Michel BA, Bloch DA, Calabrese LH, Hunder GG, Arend WP, et al. The American College of Rheumatology 1990 criteria for the classification of Henoch-Schönlein purpura. *Arthritis Rheum*. 1990;33(8):1114-21.
11. Anil M, Aksu N, Kara OD, Bal A, Anil AB, Yavaşcan O, et al. Henoch-Schönlein purpura in children from western Turkey: a retrospective analysis of 430 cases. *Turk J Pediatr*. 2009;51(5):429-36.
12. Alfredo CS, Nunes NA, Len CA, Barbosa CM, Terreri MT, Hilário MO. Henoch-Schönlein purpura: recurrence and chronicity. *J Pediatr (Rio J)*. 2007;83(2):177-80.
13. Ozen S, Ruperto N, Dillon MJ, Bagga A, Barron K, Davin JC, et al. EULAR/PreS endorsed consensus criteria for the classification of childhood vasculitides. *Ann Rheum Dis*. 2006;65(7):936-41.
14. Khader M, Ammayreh W, Issa A, Abdallat S, Momani B. Henoch-Schönlein purpura in Jordanian children. *Middle East Journal of Family Medicine*. 2008;6(1):15-6.
15. Kumar L, Singh S, Goraya JS, Uppal B, Kakkar S, Walker R, et al. Henoch-Schönlein purpura: the Chandigarh experience. *Indian Pediatr*. 1998;35(1):19-25.
16. Yang YH, Hung CF, Hsu CR, Wang LC, Chuang YH, Lin YT, et al. A nationwide survey on epidemiological characteristics of childhood Henoch-Schönlein purpura in Taiwan. *Rheumatology (Oxford)*. 2005;44(5):618-22.
17. al Harbi NN. Henoch-Schönlein syndrome in children: experience from southern part of Saudi Arabia. *East Afr Med J*. 1996;73(3):191-3.
18. Calviño MC, Llorca J, García-Porrúa C, Fernández-Iglesias JL, Rodríguez-Ledo P, González-Gay MA. Henoch-Schönlein purpura in children from

- northwestern Spain: a 20-year epidemiologic and clinical study. *Medicine (Baltimore)*. 2001;80(5):279-90.
19. Blanco R, Martínez-Taboada VM, Rodríguez-Valverde V, García-Fuentes M, González-Gay MA. Henoch-Schönlein purpura in adulthood and childhood: two different expressions of the same syndrome. *Arthritis Rheum*. 1997;40(5):859-64.
20. Yang YH, Huang MT, Lin SC, Lin YT, Tsai MJ, Chiang BL. Increased transforming growth factor-beta (TGF-beta)-secreting T cells and IgA anti-cardiolipin antibody levels during acute stage of childhood Henoch-Schönlein purpura. *Clin Exp Immunol*. 2000;122(2):285-90.
21. Allen DM, Diamond L K, Howell DA. Anaphylactoid purpura in children (Schonlein-Henoch syndrome): review with a follow-up of the renal complications. *Am J Dis Child*. 1960; 99:833-54.
22. Meadow SR, Glasgow EF, White RH, Moncrieff MW, Cameron JS, Ogg CS. Schönlein-Henoch nephritis. *Q J Med*. 1972;41(163):241-58.
23. Liu PM, Bong CN, Chen HH, Huang YC, Huang CC, Yang KD, et al. Henoch-Schönlein purpura with hemorrhagic bullae in children: report of two cases. *J Microbiol Immunol Infect*. 2004;37(6):375-8.
24. Fischer PJ, Hagge W, Hecker W. Schönlein-Henoch purpura. A clinical study of 119 patients with special reference to unusual complications. *Monatsschr Kinderheilkd*. 1990;138 (3):128-34. [Article in German]
25. Silva CAA, Campos LMMA, Liphaus BL, Kiss MHB. Henoch-Schönlein purpura in childhood and adolescence. *Rev Bras Reumatol* 2000; 40:128-36. [Article in Portuguese]
26. Lanzkowsky S, Lanzkowsky L, Lanzkowsky P. Henoch-Schönlein purpura. *Pediatr Rev*. 1992;13(4):130-7.
27. Shah D, Goraya JS, Poddar B, Parmar VR. Acute infantile hemorrhagic edema and Henoch-Schönlein purpura overlap in a child. *Pediatr Dermatol*. 2002;19(1):92-3.
28. Garty BZ, Ofer I, Finkelstein Y. Acute hemorrhagic edema of infancy. *Isr Med Assoc J*. 2002;4(3):228-9.
29. Narchi H. Risk of long term renal impairment and duration of follow up recommended for Henoch-Schonlein purpura with normal or minimal urinary findings: a systematic review. *Arch Dis Child*. 2005;90(9):916-20.
30. Tohmaz MM, Saleh SI, Al-Anezi F. Henoch-Schonlein purpura: presentation patterns in Arab children in Kuwait. *Middle East Journal of Family Medicine*. 2008;6(1):13-4.
31. Kaku Y, Nohara K, Honda S. Renal involvement in Henoch-Schönlein purpura: a multivariate analysis of prognostic factors. *Kidney Int*. 1998;53(6):1755-9.
32. Sano H, Izumida M, Shimizu H, Ogawa Y. Risk factors of renal involvement and significant proteinuria in Henoch-Schönlein purpura. *Eur J Pediatr*. 2002;161(4):196-201.
33. Rigante D, Candelli M, Federico G, Bartolozzi F, Porri MG, Stabile A. Predictive factors of renal involvement or relapsing disease in children with Henoch-Schönlein purpura. *Rheumatol Int*. 2005;25(1):45-8.
34. Shin JI, Park JM, Shin YH, Hwang DH, Kim JH, Lee JS. Predictive factors for nephritis, relapse, and significant proteinuria in childhood Henoch-Schönlein purpura. *Scand J Rheumatol*. 2006;35(1):56-60.
35. Dudley J, Afifi E, Gardner A, Tizard EJ, McGraw ME. Polymorphism of the ACE gene in Henoch-Schönlein purpura nephritis. *Pediatr Nephrol*. 2000;14(3):218-20.
36. Lim DC, Cheng LN, Wong FW. Could it be Henoch-Schonlein purpura? *Aust Fam Physician*. 2009;38(5):321-4.

37. Davin JC, Weening JJ. Diagnosis of Henoch-Schönlein purpura: renal or skin biopsy? *Pediatr Nephrol.* 2003;18(12):1201-3.
38. Cassidy JT, Petty RE. Vasculitis. In: Cassidy JT, Petty RE, editors. *Textbook of pediatric rheumatology.* 3rd ed. Philadelphia; W.B. Saunders Company; 1995. p. 365-422.
39. Evans-Jones LG, Clough JV. Thrombocytosis in Henoch-Schonlein syndrome. *Clin Lab Haematol.* 1990;12(2):137-9.
40. Trapani S, Micheli A, Grisolia F, Resti M, Chiappini E, Falcini F, et al. Henoch Schonlein purpura in childhood: epidemiological and clinical analysis of 150 cases over a 5-year period and review of literature. *Semin Arthritis Rheum.* 2005;35(3):143-53.

پوخته

نه خوشیا پرسکین هینوک- شونلاین لدهف زاروکین کورد ل باژیری دهوکی

پیشهکی و ئارمانج: پرسکین هینوک- شونلاین مشهترین نه ساخیا دهمارین خوینی یه کو توشی پیستی وگه ها و کوئه ندامی هه رسکرنی و گولچیسکا ل ناف زاروکا. ئە ڤه کولینه هاته کرن ژبو خواندنه ڤه یا لایه نی کلینیکی یی نه خوشیا پرسکین هینوک- شونلاین لدهف زاروکین کورد و ههروه سا ژبو دانا رونکره کی لسه ر لایه نی دهر دزانی یی ڤی نه خوشیی لباژیری دهوکی- باکوری عراقی.

ریکین ڤه کولینی: بو ڤی ڤه کولینی ئە و زاروکین نه خوشیا پرسکین هینوک- شونلاین لی هاتیه دهستنیشان کرن و چاره سه رکرن ل نه خوشخانا هیقی یا زاروکان هاتنه وهرگرتن دماوی دناڤه را چریا ئیکی 2009 هه تا تیرمه ها 2011ی. نه خوش هاتنه هه لسه نگاندن ژلایی ژی، رهگه ز، نیشانیی دهستیپیکی، هه بوونا نیشانیی دی یی کلینیکی، وهرزی نه ساخی و فاکته رین دی یی پیه وندی دار. و بو هه ر نه خوشه کی ئە ڤه تاقیکر نه هاتنه کرن: CBC, ESR و تاقیکرین شولی گولچیسکا و تاقیکرنا ده ستاڤا زراف. نمونه ک ژ پیستی هاته وهرگرتن ژ 11 نه خوشا.

ئه نجام: لسه رانسه ری دو سالی ڤه کولینی 51 زاروک هاتنه دهستنیشان کرن کو ئە ڤه نه خوشیه هه ی، و ژوان 23 کچ بوون و 28 کور. ژیی وان دناڤه را 10 هه یقی هه تا 15 سالی بووو تیکراییی ژیی وان 7.1 سال بوون. دهستیپیکرنا نه خوشیی پتر ل وهرزه ی زستانی و باییزی بوو. هه می نه خوشان پرسکین بدهست هه ستیار هه بوون. کولبوون و ئیشانا گه هین مهن لدهف 41 (80.4%) زاروکان هه بوو و ئیشانا زکی لدهف 24 (47%)، به لی توشبوونا گولچیسکی بتنی لدهف ئیک زاروک هه بوو.

دهرئه نجام: نه خوشیا پرسکین هینوک- شونلاین ئیک ژ نه خوشییی به ربه لاقه ده ڤ زاروکین مه به س سڤکتره و زور کیم گولچیسکان ڤه دگریت. پیئقییه چاڤدیریا دریز بو تووش بویا بهیته کرن بو تومارکرنا ئە وین دوجارکی تووش دبن.

الخلاصة

فرقرية هينوك-شونلاين لدى الأطفال الكرد في مدينة دهوك

خلفية واهداف البحث: فرقرية هينوك-شونلاين من أكثر امراض التهاب الأوعية الدموية ألتى تصيب الجلد و المفاصل و الجهاز الهضمي و الكليتين لدى الاطفال. أعد هذا البحث للاطلاع على الجوانب السريرية و الوبائية لفرقرية هينوك-شونلاين لدى الأطفال الكرد في مدينة دهوك.

طرق البحث: تمت دراسة الأطفال المصابين بفرقرية هينوك-شونلاين،الذين عولجوا في مستشفى هيفي التعليمي للفترة من شهر تموز 2009 و لغاية تموز 2011 من حيث العمر و الجنس و العلامات الأولية والعلامات الأخرى للمرض، و العوامل المؤهبة و الفترة التي ظهر فيها المرض.الفحوصات المختبرية التي أجريت شملت فحص الدم العام و معدل ترسيب الكريات الحمر و وظائف الكلية وفحص الإدرار العام، وأخذت خزعة من الجلد في احد عشر مريضا. تمت متابعة المرضى لفترة ستة اشهر بأجراء فحص الادرار العام ووظائف الكلية لتسجيل حالات اصابة الكلية.

النتائج: تم تشخيص 51 مريضا" خلال سنتين، منهم 28 ذكرا و 23 أنثى. تراوحت أعمار المرضى بين عشرة اشهر و خمسة عشر سنة، و معدل عمر المرضى كان 7.1 سنة و نسبة الذكور إلى الإناث كان 1:1.2 . معظم المرضى تم تشخيصهم في فصلي الشتاء والخريف. جميع المرضى كان لديهم فرقرية الجلد. التهاب او آلام المفاصل الكبيرة سجل في 41 مريضا"(80.4%)، آلام البطن في 24 (47%) مريضا" و إصابة الكلية في حالة واحدة. حصل شفاء تام في جميع المرضى. بعد ستة اشهر من متابعة المرضى لم تسجل اية اصابة في الكلية.

الاستنتاجات: : الخصائص الوبائية والاحصائية لفرقرية هينوك-شونلاين في الاطفال الكرد تشبه ما هو موجود في باقي الدول ولكنها تتميز بانها اقل شدة مع اصابات قليلة جدا في الكلية.

CROHN'S DISEASE IN AN INFANT PRESENTED WITH PERFORATION OF THE COLON: A CASE REPORT

NADIR A. GARJEES, MBChB, FIBMS*

RAFIL T. YAQO, MBChB, FIBMS**

MUHAMMED H. ALDABAKH, MBChB, FIBMS***

Submitted 2 Jul 2011; accepted 26 Feb 2012

SUMMARY

Infantile Crohn's disease is very rare and has been described as a severe illness with poor prognosis. We report the clinical, hematological, radiological and pathological findings of an 11 month-old boy with infantile colonic Crohn's disease who presented with repeated vomiting, fever and pallor. Chest X ray showed an air under the diaphragm. On laparotomy there was perforation of the colon. Although rare, Crohn's disease should be considered in the differential diagnosis of any child with severe gastrointestinal symptoms.

Duhok Med J 2012;6(1): 54-58.

Key words: Chronic inflammatory bowel disease, Crohn's disease, Infancy, Intestinal perforation

Crohn's disease, an idiopathic chronic inflammatory disorder of the bowel, involves any region of the alimentary tract from the mouth to the anus. About 70% of the patients has terminal ileitis alone and in 10%-15% there is only colonic involvement.¹ All age groups are affected but it is most commonly diagnosed between the ages of 15-35. It rarely occurs in children below 10 years and it is extremely rare in infants less than 1 year of age.² We report an 11 month-old infant who presented with intestinal perforation due to Crohn's disease.

CASE PRESENTATION

An 11-month-old boy was admitted to the emergency department at Heevi Pediatric Teaching Hospital in Duhok governorate suffering from repeated vomiting and fever. On physical examination the child was dehydrated and pale and had rigid abdomen. Complete blood test was done which revealed a hemoglobin

concentration of 5 gm/dl and total white blood cell count of 37,600/mm.³ Plain abdominal x ray showed air under the diaphragm (Figure 1). The patient received blood transfusion. Written informed consent was obtained from the patient's guardian. Then exploratory laparotomy was done which showed multiple colonic perforations. The bowel walls were inflamed and matted together by thin fibrinous adhesion. Colostomy done at the site of perforation and biopsy was taken. Postoperatively, the patient started to regain his bowel function, became better generally well and started oral feeding. At the third postoperative day, the general conditions of the patient deteriorated, the colostomy stopped functioning and the site of drain started discharging fecal materials. Another exploratory laparotomy was done which showed multiple perforations at the left descending and sigmoid colon distal to site of previous colostomy. Peritoneal irrigation was performed followed by resection of the

* Lecturer in Pediatric Department, Faculty of Medical Sciences, School of Medicine, University of Duhok

** Lecturer in Pathology Department, Faculty of Medical Sciences, School of Medicine, University of Duhok

*** Lecturer in Pediatric Surgical Department, Faculty of Medical Sciences, School of Medicine, University of Duhok

Correspondence author: Nadir A. Garjees. Cell phone number 009647504820882. Email: nadir_brivkani@uod.ac

perforated segment of the colon. The removed part of the colon was sent for histopathological investigation. Grossly, the segment showed mucosal surface with multiple ulcerations and normal skipping areas giving rise to a cobblestone appearances. The wall is thickened with adhesion of creeping of pericolic fat (Figure 2). Microscopically sections of the large bowel wall showed variable depth ulceration, necrosis with chronic inflammatory cells (plasma cells, lymphocytes, histocytes and few giant cells) and non-caseating vague granulomas near the blood vessels with pericolitis (Figure 3 and 4).

DISCUSSION

Crohn's disease has rarely been observed in infancy.³ Between 1990 and 1998, three siblings and their first degree cousin were found to have Crohn's disease and their respective ages on diagnosis were 3 weeks, 2 weeks, 3 months and 2 months.⁴ In these cases, the inflammatory process involved extensive anorectal and colon involvement with sparing of the small bowel. There were severe perianal disease consisting of ulceration, mucosal and skin tags, fistulae and fissures. In our patient, there was also

involvement of colon with sparing of small bowel. The main presentation of our patient was chronic constipation followed by intestinal perforation. The first known infant affected by Crohn's disease was reported by Koop et al in 1947.⁵ Miller and Larson⁶ reported 12 cases of Crohn's disease in the newborn period. In 8 of the cases reviewed, the disease involved the small bowel, whereas in 3 the disease involved the right colon and terminal ileum; one patient had only colonic involvement. Seven of 12 patients reviewed by Miller and Larson died after surgery. The patients reported by Koop et al and Miller and Larson had a prodromal period of diarrhea lasting from 2 days to several weeks, followed by signs of intestinal obstruction. None of these had any perianal involvement. Mezot et al⁷ reported a 4-week-old infant with signs and symptoms suggesting Crohn's disease that initially associated with central nervous system thrombosis and later, at the age of 6 months, with severe perianal inflammations. In conclusion, although rare, Crohn's disease in infancy should be considered in the differential diagnosis of any infant with severe chronic gastrointestinal manifestation.



Figure 1. Chest X ray shows air under the diaphragm

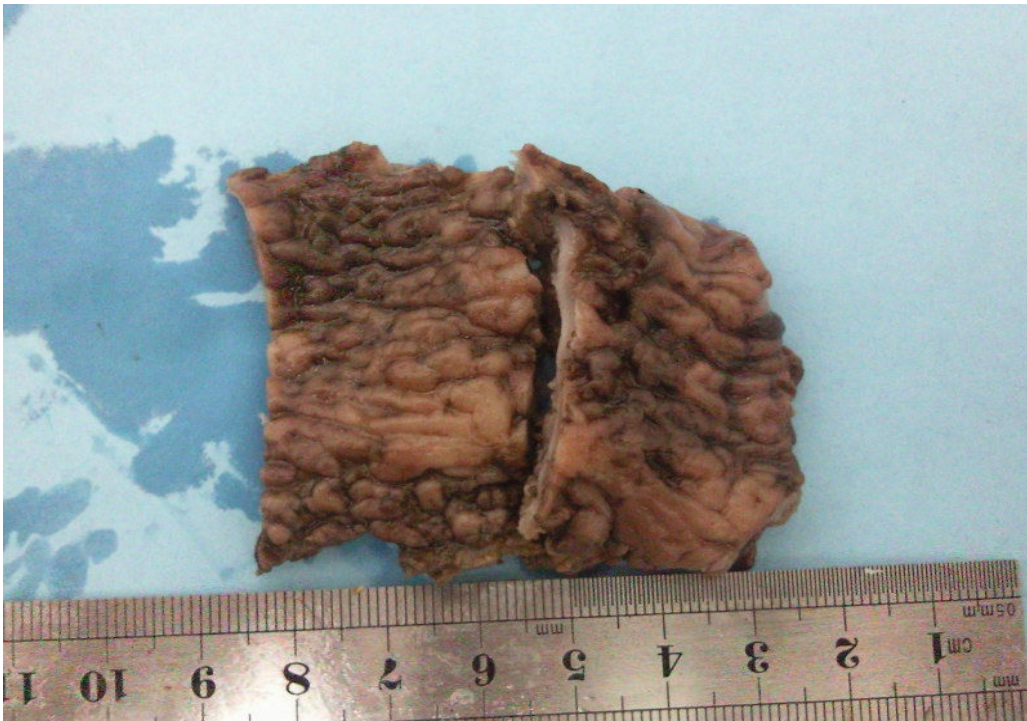


Figure 2. Large bowel segment reveal cobblestone appearances with presence of skipping lesion

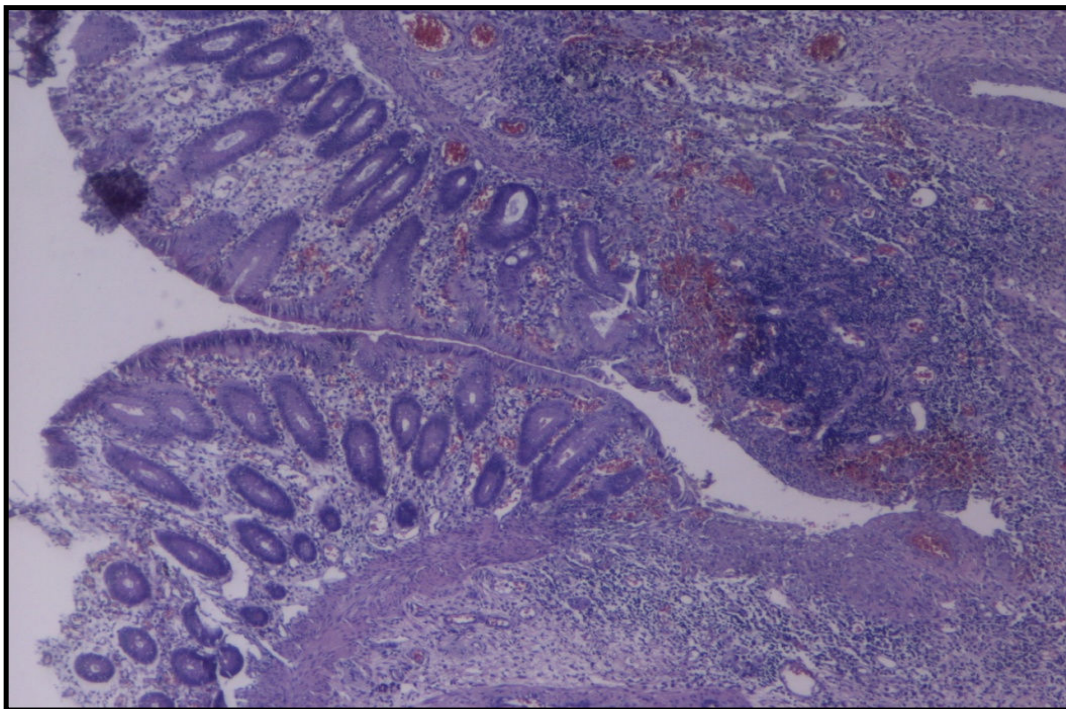


Figure 3. Ulcer that reach the sub mucosa with vague granuloma at the base and normal edematous mucosa at edges of the ulcer

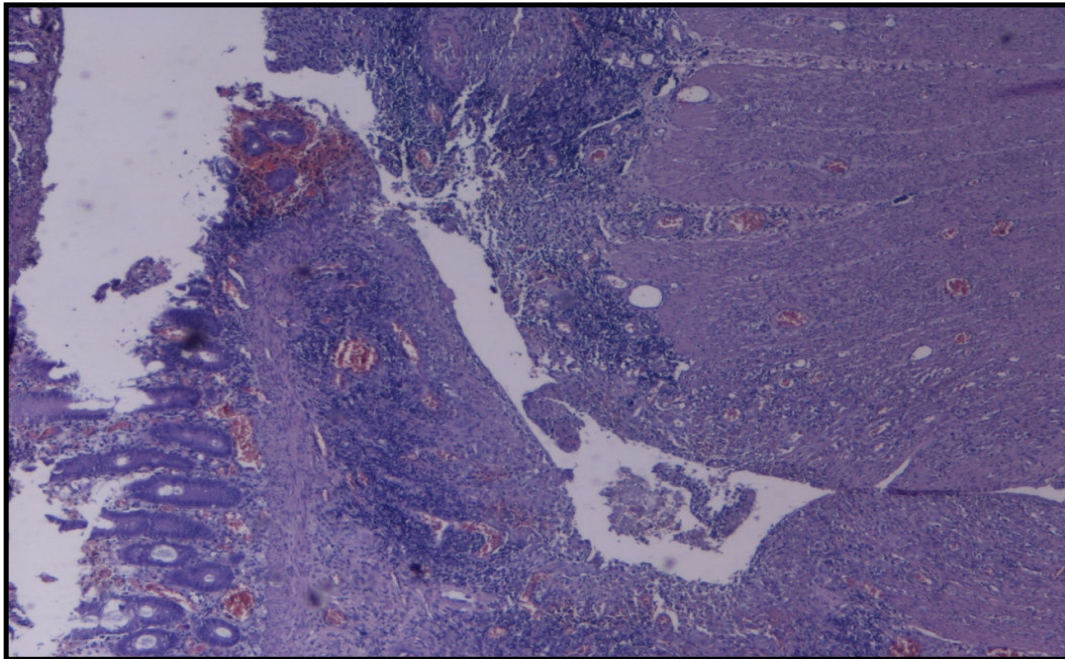


Figure 4. Fissure through the muscularis propria

REFERENCES

1. Buamgart DC, Sandborn WJ. Inflammatory bowel disease: clinical aspect and established and evolving therapies. *Lancet*. 2007; 369(9573):1641-57.
2. Vargas JH. Medical management of Crohn's disease in childhood. *Semin Pediatr Surg*. 1994; (1):15-8.
3. Gryboski JD. Crohn's disease in children 10 years old and younger: comparison with ulcerative colitis. *J Pediatr Gastroenterol Nutr*. 1994;18(2):174-82.
4. Cohen Z, Weizman Z, Kurtzbar E, Newman N, Kapuller V, Maor, et al. a report of four cases in one family. *J Pediatr Gastroenterol Nutr*. 2000;30(4):461-3.
5. Koop CE, Perlingiero JG, Weiss W. Cicatrizing enterocolitis in a newborn infant. *Am J Med Sci*. 1947;124(1):27-32.
6. Miller RC, Larsen E. Regional enteritis in early infancy. *Am J Dis Child*. 1971;123(4):301-11.
7. Mezoff AG, Cohen MB, Maisel SK, Farrell MK. Crohn's disease in an infant with central nervous system thrombosis and protein-losing enteropathy. *J Pediatr*. 1990; 117(3):436-9.

پوخته

نیشا کرونی ل زاروکی دگهل هه بونا کونی ل قولونی (ریفیکا ستویر): دۆزا توماری

نساخیا کرونی ل سافایان زورا دهگمه نه و هاتییه رافه کرن وهك نیشهكا گران و بهربینییهكا کیم. مه تومارکرب ریکا دیتنت کلین و هیماتولوجی (بکار ئینانا خینی) و تیشکی و بایوپسی (وه رگرتنا نمونه ی ل لهشی و تاقیکرنا وی) ل زاروکی کور 11 هه یفی کو دنالی ب سدهما نساخیا کرونی یا قولونی کو پیقه دیاربو قهره شبانهكا بهردهوام و تا و زه رایبی. ب تیشکا X یا سینگی هاته دیارکرن کو ههوا ل بن پهردهیا نافبرئ دا ههیه و دقه کرنا زکی دا کونیونا قولونی هه بو هه رچه ند یا کیم بوو. ئەف نساخیه دفتت بهیتته بهر چاف کرن ل هه زاروکی کونی ئه وین دنالین ب سدهما نیشانین ئیشین ئورک و ریفیکان.

الخلاصة

داء الكرون في الاطفال يتمثل بوجود ثقب في القولون: تقرير الحالة

مرض الكرون في الاطفال نادر الحدوث ويوصف بشدة المرض وصعوبة تشخيصه. حيث يأخذ الحالة السريرية واجراء فحوصات الدم والأشعة واخذ عينة من القولون للأطفال التي اعمارهم 11 شهر والمصابون بهذا المرض والذي لديهم التقبوء والحمى والشحوب، عند اجراء اشعة الصدر يلاحظ وجود الهواء تحت الحجاب الحاجز وجراحيا يلاحظ ثقب في القولون مع انه نادر الحدوث. مرض يجب أن يؤخذ بنظر الاعتبار في التشخيص التفاضلي لأي طفل بالأعراض المعوية الحادة.

AMIODARONE- INDUCED THYROIDITIS IN A POST-CARDIAC TRANSPLANT PATIENT – CASE REPORT

HATEM AL-FARHAN, MBBS, MRCP(UK)*

AFTAB A. SIDDIQUI, MBBS, FCPS, MRCP (UK), FRCP(Edin), FRCP (Ireland)*

YASSER W. SHAREF, MBChB, MRCP (Ireland)*

ADIL F. AL LAWATI, MD, MRCP (UK)*

Submitted 27 Jan 2012; accepted 5 Jun 2012

SUMMARY

Amiodarone is a class III antiarrhythmic medication that is widely used for the treatment of various arrhythmias and is useful in non-ischaemic dilated cardiomyopathy. It is well known that amiodarone can alter the biochemical status of the thyroid gland. Amiodarone-induced thyroiditis is more common than amiodarone-induced hypothyroidism. It is estimated that 23% of patients receiving amiodarone may develop amiodarone-induced thyroiditis. We are reporting a case of a young lady with familial dilated cardiomyopathy who was on amiodarone that was stopped in July 2007, yet she developed clinical signs and biochemical evidence of amiodarone-induced thyroiditis few months after her cardiac transplantation despite stopping amiodarone. She was managed conservatively jointly with the endocrinologist and her thyroid function test returned back to normal eventually.

Duhok Med J 2012;6(1): 59-63.

Key words: Amiodarone, Thyroiditis, Dilated cardiomyopathy, Cardiac transplant

Amiodarone is an effective antiarrhythmic medication that is used in treatment of various arrhythmias and it is also useful in non-ischaemic dilated cardiomyopathy. It is a benzofuran-derived, iodine-rich drug with many structural similarity to thyroxine (T₄). Amiodarone contains around 37% iodine by weight. Every 200-mg tablet is estimated to have about 75 mg of organic iodide, 8-17% of which is released as free iodide. Standard maintenance treatment with 200 mg amiodarone can provide more than 100 times the usual daily iodine requirement. It is highly lipid-soluble and is concentrated in the adipose tissue, lung, muscle, liver, and thyroid gland. It may cause severe side effects, including thyroiditis; hypothyroidism and thyrotoxicosis. This may present even after discontinuation of the drug as the elimination half-life of amiodarone is

variable, ranging from 50-100 days; total body iodine stores remain increased for up to 9 months after discontinuation of the drug.

CASE PRESENTATION

A 20-year old Omani lady who has been diagnosed to have “familial dilated cardiomyopathy” complicated with congestive heart failure for three years with intracardiac defibrillator (ICD) being inserted in 2005. She was on warfarin, spironolactone, furosemide, valsartan, carvedilol, amiodarone and ranitidine. She was first admitted in Sultan Qaboos University Hospital (SQUH) in January 2007 with community acquired pneumonia (CAP) complicated by decompensated heart failure status. She was found to have BP of 80/50 mmHg, tachycardia of 110 beats per minute,

* Department of Medicine, Sultan Qaboos University Hospital, Muscat, Sultanate of Oman

Correspondence author: Dr. Aftab A. Siddiqui, Consultant in Internal Medicine, Department of Medicine, Sultan Qaboos University Hospital, POBOX 35, AlKhodh 123, Muscat, Sultanate of Oman; E-mail: aftabsidpk@hotmail.com, Fax: +968 24141198

displaced apex, mitral regurgitation (MR) and tricuspid regurgitation (TR) murmurs, congestive hepatomegaly and lower limbs edema. The laboratory investigations on admission were as shown in table 1.

Table 1. Baseline investigations as on first admission

Test	Result
Haemoglobin	13.9 g/dl
WCC	5.8 x 10 ⁹ /L
Neutrophils	3.0 x 10 ⁹ /L
Platelet	187000 /L
Serum sodium	128 mmol/L
Serum potassium	4.1 mmol/L
Serum chloride	97 mmol/L
Serum bicarbonate	23 mmol/L
Serum urea	6.1 mmol/L
Serum creatinine	58 umol/L
eGFR	> 90 ml/min/1.73 m ²

Her chest X-ray showed cardiomegaly and pulmonary congestion along with lobar pneumonia in right lower lobe. Her echocardiography (ECHO) showed grossly dilated 4 heart chambers with severe global hypokinesia. Her LVEF was 28% and moderate MR and TR. Her thyroid function tests (TFT) on admission were within the reference ranges.

She was treated with anti-heart failure medications and antibiotics, she made good recovery from her CAP and her heart failure was reasonably controlled. She was discharged on 07/07/2007 in a stable condition with a follow up in cardiology clinic. Her medications included all the

drugs mentioned above apart from amiodarone which was stopped.

The patient underwent cardiac transplantation in China in August 2007. In her follow up at SQUH in November 2007 she was asymptomatic and free from signs of cardiac failure. However, she was found to have diffusely enlarged goiter with no tenderness and no overlying bruit or local lymphadenopathy. Nevertheless, her laboratory investigations including full blood count, renal and liver functions were all within the reference ranges.

Her repeated ECHO showed mild left ventricular hypertrophy with mild hypokinesia of the anterior wall. Her LVEF was 60% with only trivial MR and TR. Post transplantation, she was kept on captopril 25 mg TID, sirolimus 2 mg OD, mycophenolate 750 mg BiD and carvedilol 12.5 mg BD. Her TFT showed TSH 0.02 mU/L, FT3: 8 pmol/L and FT4 27 pmol/L, pattern of low TSH with slightly raised FT3 and FT4. Thyroid antibodies were negative. Thyroid uptake scan was done and showed global low uptake of 0.1% suggestive of thyroiditis (Figure 1). However, the patient remained clinically euthyroid. She was reviewed by endocrinologist who advised close monitoring and not to start any anti-thyroid medications like thioamides. The patient had serial TFTs which returned back to normal ranges in July 2008 and she maintained these values to date (Table 2). The patient was euthyroid throughout this period.

Table 2. Thyroid function tests on admission and during follow up

TFT	6/11/2007	15/1/2008	29/7/2008	6/1/2009	7/7/2009
FT3 (3.8-6 pmol/l)	7.8	9.5	5.1	5.1	4.6
FT4 (7.9-14.4 pmol)	18.2	27.0	8.1	9.7	8.1
TSH (0.34-5.6 mIU/l)	0.02	0.02	1.51	1.31	1.08

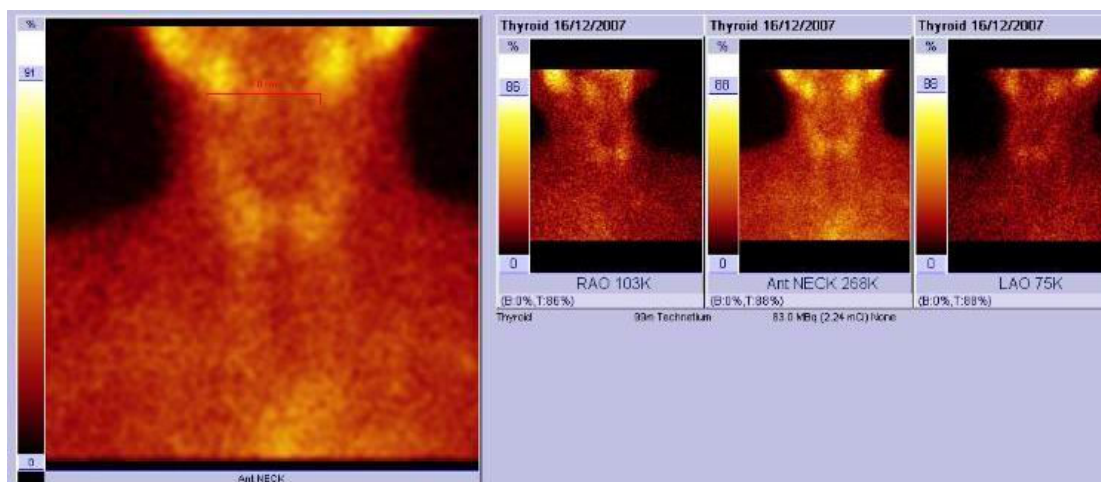


Figure 1. Thyroid uptake scan showing low uptake of 0.1%

DISCUSSION

Amiodarone is a class III antiarrhythmic medication that is widely used in the treatment of various arrhythmias and is useful in non-ischaemic dilated cardiomyopathy (NIDCM).¹ It is well known that amiodarone can alter the biochemical status of the thyroid gland. It is estimated that 23% of patients receiving amiodarone develop amiodarone-induced thyroiditis (AIT).²

Amiodarone induced thyroiditis can be classified into two types according to their mechanism. In type 1 which is the latent disorder, there is synthesis and release of excessive thyroid hormones whereas in type 2 which is more common, there is increase in the release of thyroid hormones due to the destructive effect of amiodarone on the thyroid gland.² Thyroid antibodies, iodine uptake scan and doppler studies can be used to differentiate the two forms. Combination of both types was reported in some patients and differentiation of the two types is not always easy.²

Interestingly, AIT can present despite discontinuation of amiodarone therapy. Patients who were on amiodarone before heart transplantation are prone to AIT for a year after discontinuing amiodarone. Careful monitoring of thyroid function is recommended for those patients.³

Patients with AIT are known to be prone to major adverse cardiovascular event (MACE). In a study done in Hong Kong enrolling 354 patients defined MACE as cardiovascular mortality, myocardial infarction, stroke and heart failure, or ventricular arrhythmias that required hospitalization. They concluded that AIT is associated with a 2.7-fold increased risk of MACE.⁴

Although amiodarone-associated thyroid dysfunction is usually a mild clinical condition that can subside spontaneously in approximately 20% of patients.⁵ However, it can be severe, life threatening, and even fatal. However, the long term prognosis for AIT is usually good.⁶

In conclusion, Amiodarone-associated thyroid dysfunction can present despite discontinuation of amiodarone therapy as this drug has a very long elimination half life and this is more in those with slow acylators. It is important for physicians who regularly prescribe amiodarone to be aware of its consequences on the thyroid function especially in long-term users. Thyroid hormone levels and antibodies should be done as a baseline and repeated every six months or earlier if clinical features of thyroid dysfunction appear in patients on amiodarone.

REFERENCES

1. Strickkberger SA, Hummel JD, Bartlett TG, Frumin HI, Schuger CD, Beau SL, et al. Amiodarone versus implantable cardioverter-defibrillator: randomized trial in patients with nonischemic dilated cardiomyopathy and asymptomatic nonsustained ventricular tachycardia-AMIOVIRT. *J Am Coll Cardiol.* 2003;41(10):1707-12.
2. Pearce EN, Farwell AP, Braverman LE. Thyroiditis. *New Engl J Med.* 2003; 348: 2646-55.
3. Siccama R, Balk AH, de Herder WW, van Domburg R, Vantrimpont P, van Gelder T. Amiodarone therapy before heart transplantation as a predictor of thyroid dysfunction after transplantation. *J Heart Lung Transplant.* 2003; 22(8): 857-61.
4. Yiu KH, Jim MH, Siu CW, Lee CH, Yuen M, Mok M, et al. Amiodarone-induced thyrotoxicosis is a predictor of adverse cardiovascular outcome. *J Clin Endocrinol Metab.* 2009;94(1): 109-14.
5. Eaton SE, Euinton HA, Newman CM, Weetman AP, Bennet WM. Clinical experience of amiodarone induced thyrotoxicosis over a 3-year period; role of colour-flow Doppler sonography. *Clin Endocrin.* 2002; 56(1): 33-8.
6. Vorperian VR, Havighurst TC, Miller S, January CT. Adverse effects of low dose amiodarone: a meta-analysis. *J Am Coll Cardiol.* 1997;30(3):791-8.

پوخته

نه خوشيا تايرودايتس ژ نه گيرئ ب كارئينانا نه مايوداروني پشتي نه خوشيا چاندنا دلي – راپورتكرنا حاله ته كي

نه مايودارون ئيك ژ دهرمانين د هيت ه ب كارئينانا بو ريځكستنا ليدانا دلي ههروهسا بو نه خوشيا (Non-ischemic dilated cardiomyopathy). و دهيت ه پيشبينكرن كو 23٪ ژ نه خوشين نه ئي دهرمانى ب كاريين توشى ههودانا تايرودى بن. نه ئه راپورتا نه خوشه كا ئافره ته كو نه خوشيا (Familial dilated cardiomyopathy) كول ته مموزا 2007 نه دهرمانه ب كار نه ئيناي به لي هه ر توشى ههودانا تايرودى بوى چهند هه يقا پشتي نشته رگه ريا چاندنا دلي. نه د نه خوشه هاته چاره سه ركين ب ريځكين ئاساي وهك ب كارئينانا نه نتي بايوتيك و دهرمانين دژى وهستاندا دلي و تافيكركنا كاري تايرودى هاته زفراندان بو حاله تي نورمال.

الخلاصة

التهاب الغدة الدرقية نتيجة استعمال الامايودارون بعد اجراء عملية زرع القلب – اشهار حالة

الامايودارون هو أحد الادوية التي تستعمل في علاج عدم انتظام ضربات القلب كما هو مفيد في علاج مرض (Non-ischemic dilated cardiomyopathy). و من المعروف بأن هذا الدواء يؤدي الى خلل في وظائف الغدة الدرقية. و يذكر بأن التهاب الغدة الدرقية نتيجة استعمال هذا الدواء هو أكثر شيوعا من نقص هورمون الغدة الدرقية نتيجة استعمال الدواء. و قد قمنا برصد حالة شابة مصابة ب (Familial dilated cardiomyopathy) و التي كانت تستعمل الدواء الى حين توقف الدواء في تموز 2007. رغم عدم استعمال الدواء فقد ظهرت لدى المريضة العلامات السريرية و البايوكيمياوية لالتهاب الغدة الدرقية نتيجة استعمال هذا الدواء عدة أشهر بعد اجراء عملية زرع القلب. تم علاج المريضة تحفظيا حيث تم ارجاع وظائف الغدة الدرقية الى حالتها الطبيعية.